### Many transcription factors recognize DNA shape

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## Most disease associated mutations are outside coding regions

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<u>Approach</u>: Map variants to correct pathways by predicting enhancers and their target genes. Score variants for changes in binding affinity.

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# MotifDiverge quantifies loss/gain of TF binding sites

Statistical model for TFBS evolution with turnover

Seq A Seq A  $P(N_m = n_m) =$ Bernoulli trial  $(n_A - n_B)$   $Bernoulli trial <math>(n_A - n_B)$   $H Hits <math>(m_A)$   $H Hits <math>(m_B)$  $H Hits <math>(m_B)$ 

6) 
$$\begin{cases} \sum_{j=0}^{k_x - k_y} P_s(N_1 = n_{xy} - j)Bin(N_2 = j) & \text{for } k_x \ge k_y \\ \sum_{k_y - k_x} P_s(N_1 = n_{xy} + j)Bin(N_2 = j) & \text{for } k_x < k_y, \end{cases}$$

P-value for net change in binding
One or many TFs
Alignment-free
Evolutionary model
Motif specific

#### **Predicts change of function**

associated variant



#### Detects loss/gain of function mutations with high accuracy

- Better than conservation scores
- In vivo and MPRAs in cell lines

Ritter et al. (2010) Kostka et al. (2015)



Training Data Active enhancers Expressed genes Hi-C interactions Functional Genomics



**Reveals distinct genomic signature of looping DNA** 

- Heterochromatin on loop
- Cohesin within 6Kb of enhancer and promoter but not mid-loop
- TFs bound with CTCF
   improve predictions

### **Summary and Challenges**

 Machine-learning on biologically validated enhancers identifies non-coding variants most likely to affect gene regulation <u>and</u> the targeted genes.

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  Functional variants outside enhancers



<u>Hypothesis 2</u>: Non-coding variants alter binding sites of structural proteins and chromatin modifiers. Reveals distinct genomic signature of looping DNA

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#### <u>Hypothesis 2</u>: Non-coding variants alter binding sites of structural proteins and chromatin modifiers.

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- <u>Hypothesis 2</u>: Non-coding variants alter binding sites of structural proteins and chromatin modifiers.
- <u>Approach</u>: CRISPR edit sites identified by TargetFinder, then test chromatin and expression.

Reveals distinct genomic signature of looping DNA

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  - Enhancer variants outside sequence motifs

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For a typical ENCODE TF 23% of the top 2000 ChIPseq peaks have no sequence motif (range = 1%-63%)

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<u>Approach</u>: Algorithm to learn **shape motifs** de novo for all ENCODE TFs, predict shape motif hits in ChIPseq peaks, compare to sequence motifs

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  - Gibbs sampling with scores ~  $exp(\sum D_{ij})$
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- 4. <u>Enrichment test</u>: Hypergeometric p-value.

#### Shape motifs are common



#### Shape complements sequence motifs

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- Many peaks have sequence and shape motifs.
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Underlying sequence

FactorBook sequence motif

#### Cfos ProT motif in K562

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#### Shape motifs can flank sequence motifs



Underlying sequence

FactorBook sequence motif is <u>3bp</u> upstream

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Cfos Roll motif in K562





Underlying sequence

FactorBook sequence motif is <u>30bp</u> away

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- Shape motifs can flank sequence motifs
- Shape motifs can differ between TFs with similar sequence motifs and/or the same protein fold.



Fosl1 has a HelT motif



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  - Co-factors and complexes
  - Weak ChIP-seq peaks
- Role of shape in ectopic binding of TFs when cofactors are absent [Luna-Zurita et al. 2016]
- Evolutionary modeling of DNA shape
  - Conservation of shape without sequence
  - Scoring SNPs for effects on shape motifs

#### Collaborators

EnhancerFinder Tony Capra Gen Haliburton DNA Shape Hassan Samee

TargetFinder Rebecca Truty Sean Whalen MotifDiverge Dennis Kostka Functional Assays Hane Ryu Alex Pollen Nadav Ahituv Arnold Kriegstein

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