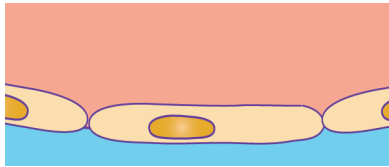


June 29-July 1, 2015  
ENCODE 2015:  
Research Application and Users Meeting

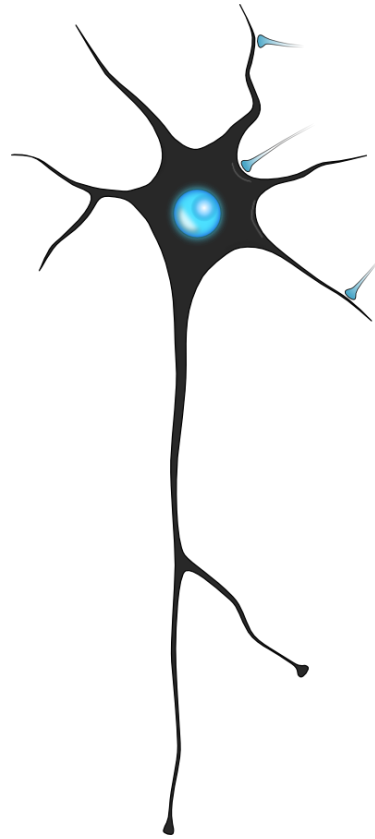
**Selection and function of signal-dependent  
enhancers:  
The Macrophage as a Case Study**

Christopher K Glass MD, PhD  
Department of Cellular and Molecular Medicine  
Department of Medicine  
University of California, San Diego

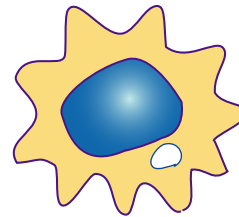
# Different cell types arise from differential transcription of the same genome



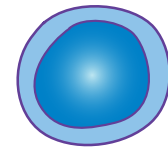
Endothelial cells



Neurons



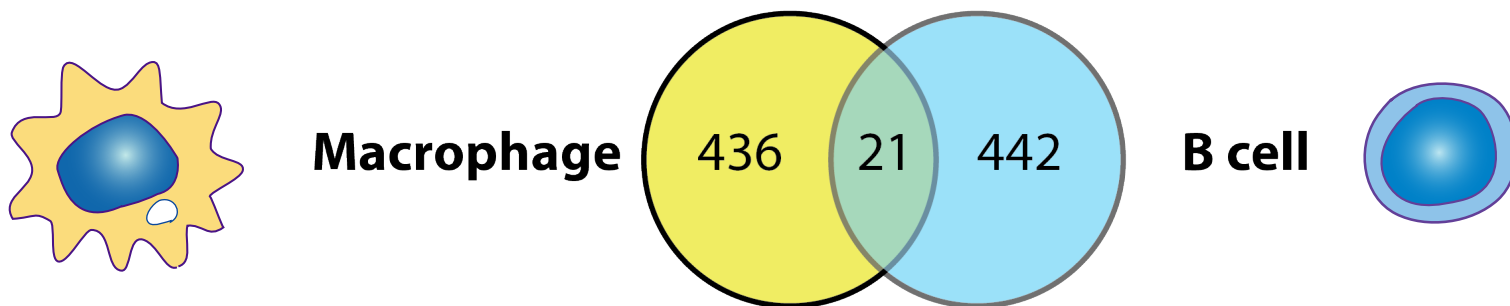
Macrophages



B cells

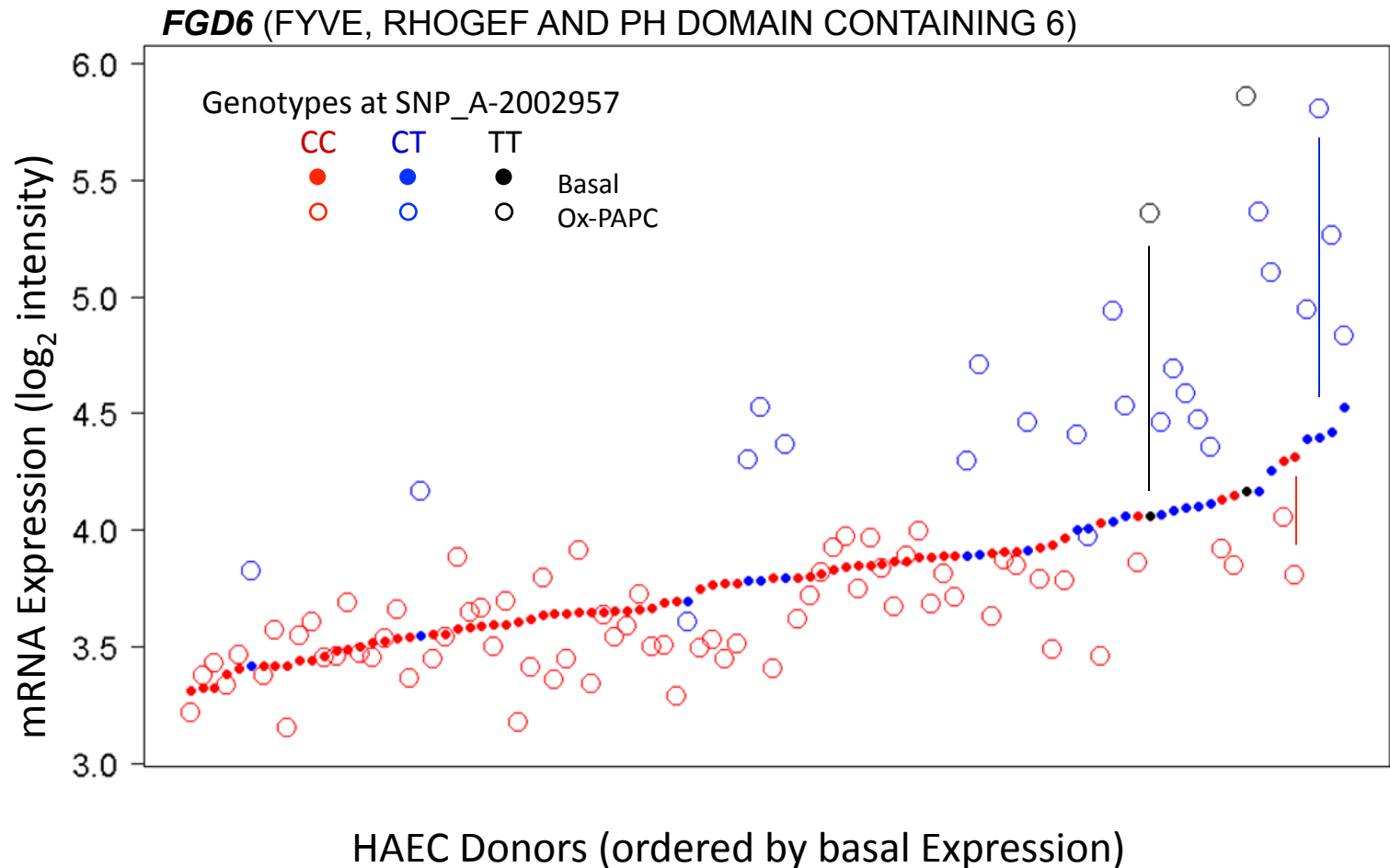
# Transcriptional responses to the same signal can be cell type-specific

mRNAs increased > 4-fold 1h after TLR4 ligation



	Macrophage	B cell
GO term	p value	p value
Immune response	1e-36	> 0.05
Chemotaxis	1e-12	> 0.05
Protein folding	> 0.05	1e-10
RNA processing	> 0.05	1e-9
DNA replication	> 0.05	8e-3

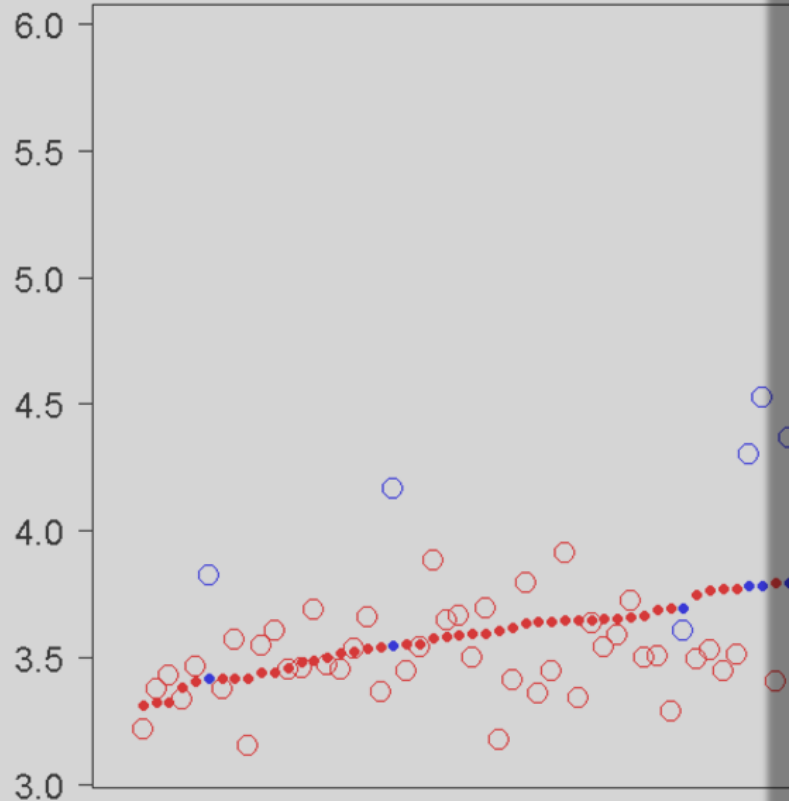
# Signal-dependent responses of the same cell type can vary among individuals



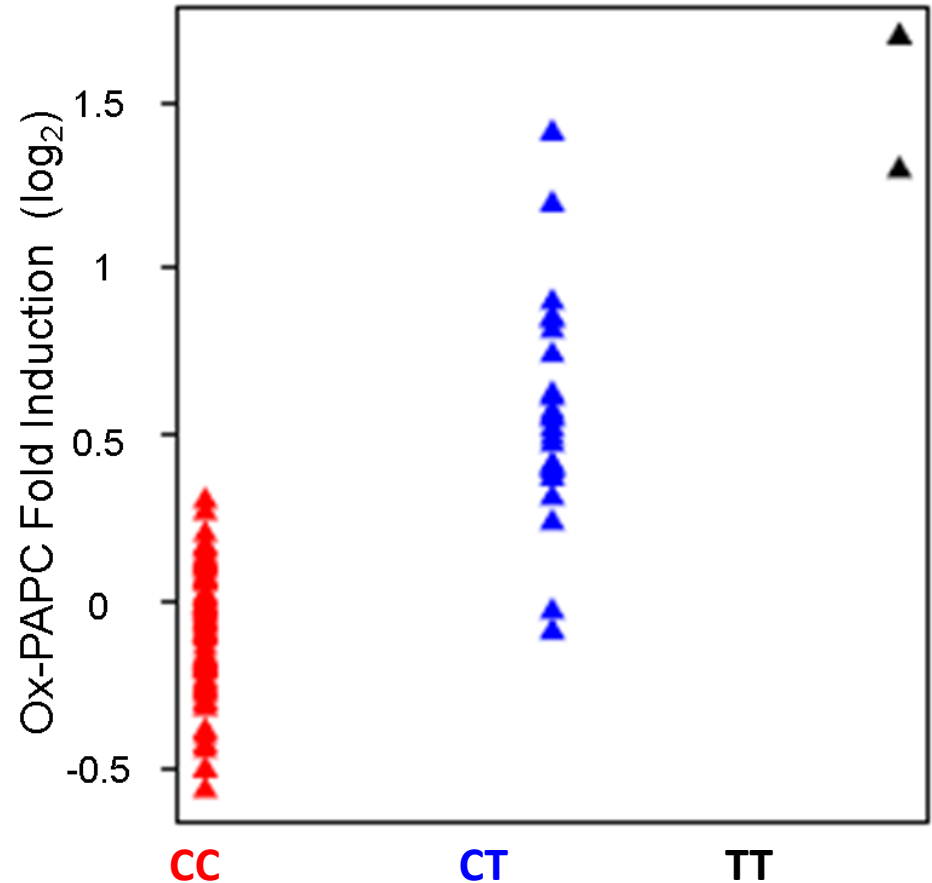


# The FGD6 response to Ox-PAPC is associated to a local DNA variant

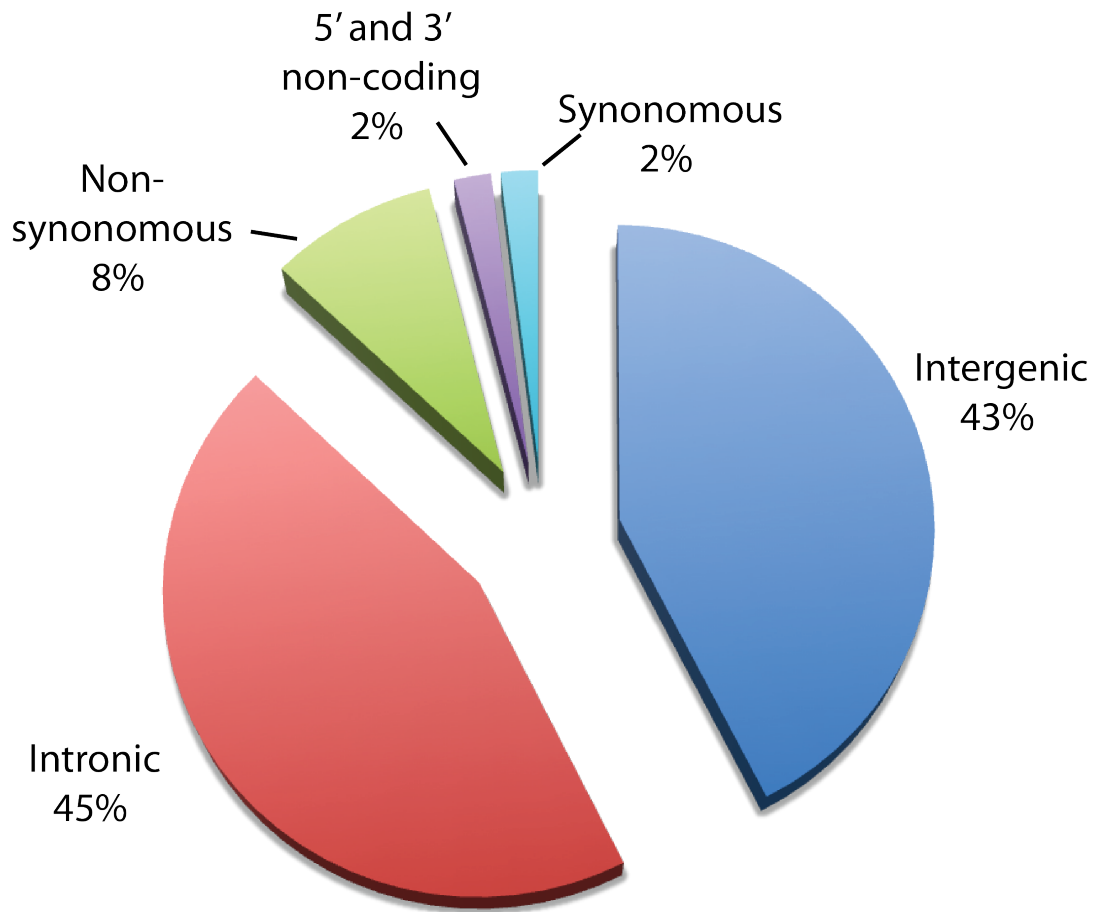
219901\_at FGD6 by



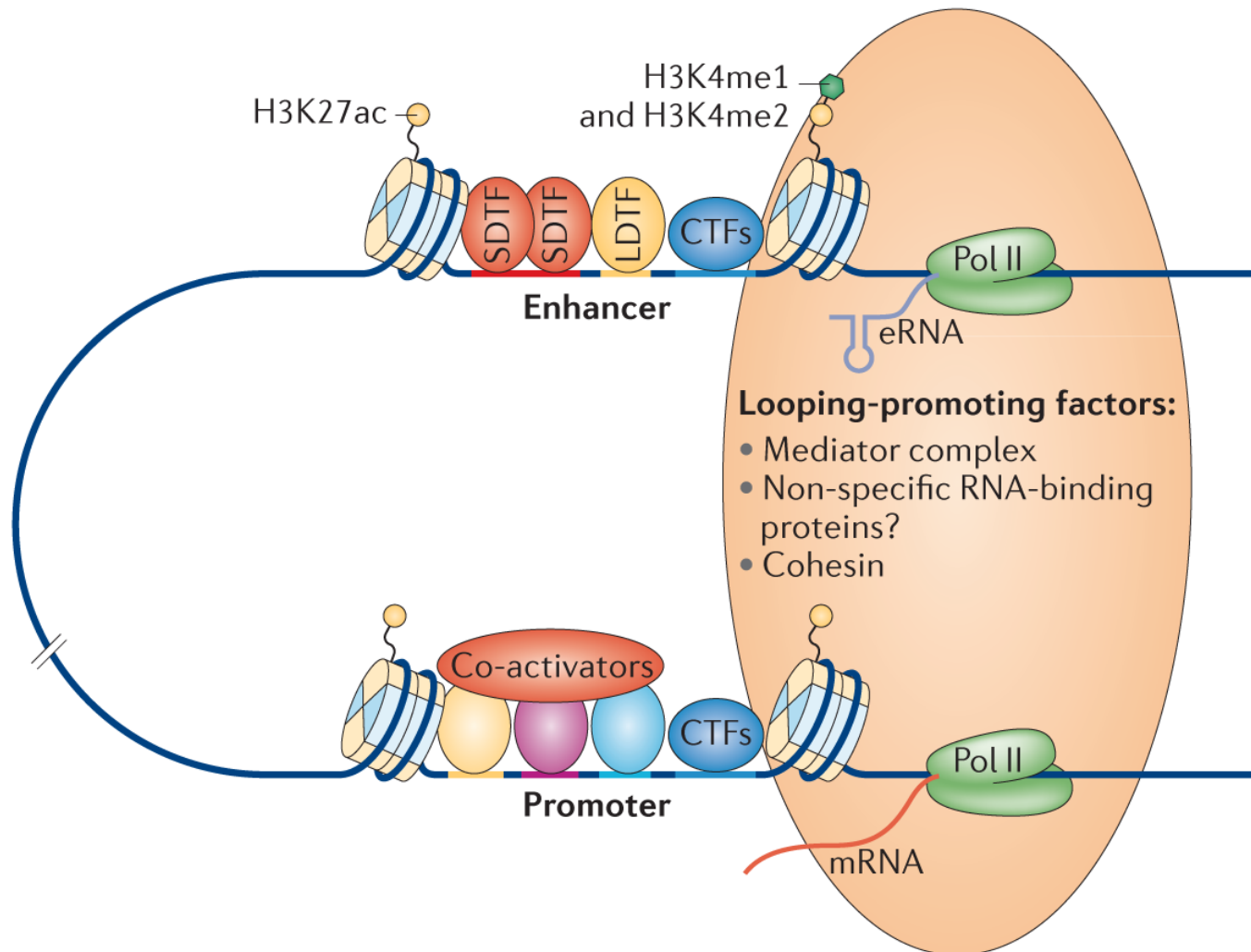
### Ox-PAPC Response of FGD6



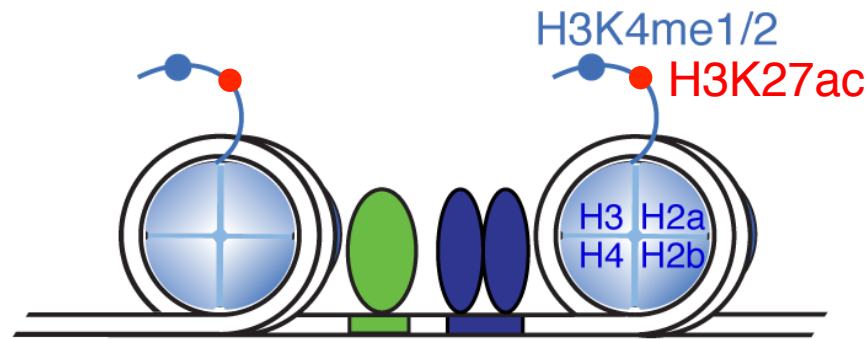
# Most GWAS disease/trait-associated SNPs reside in non-coding regions of the genome



# Enhancer/promoter interactions establish cell-specific and signal-dependent gene expression

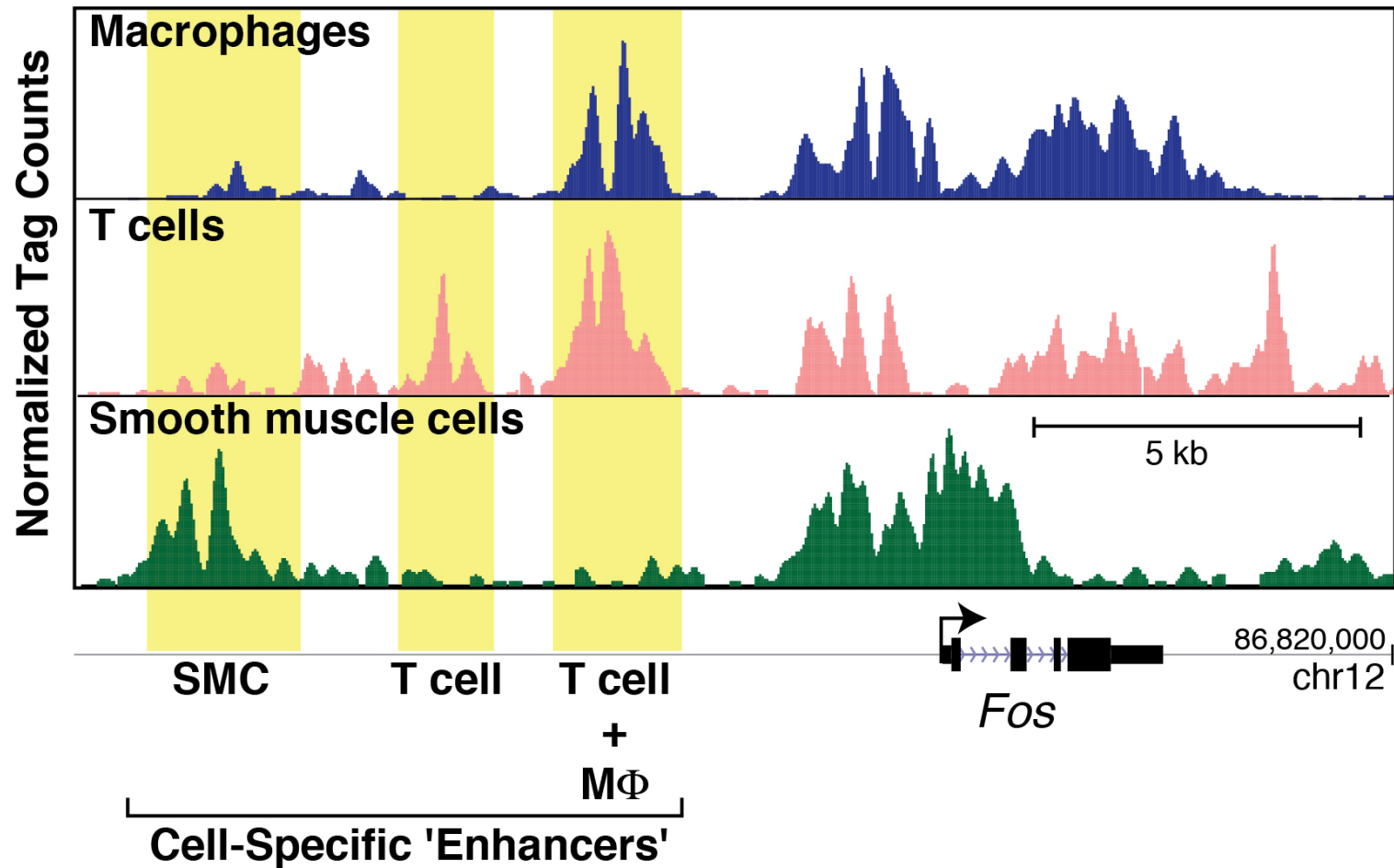


# Enhancers are major determinants of cell-specific and signal-dependent gene expression

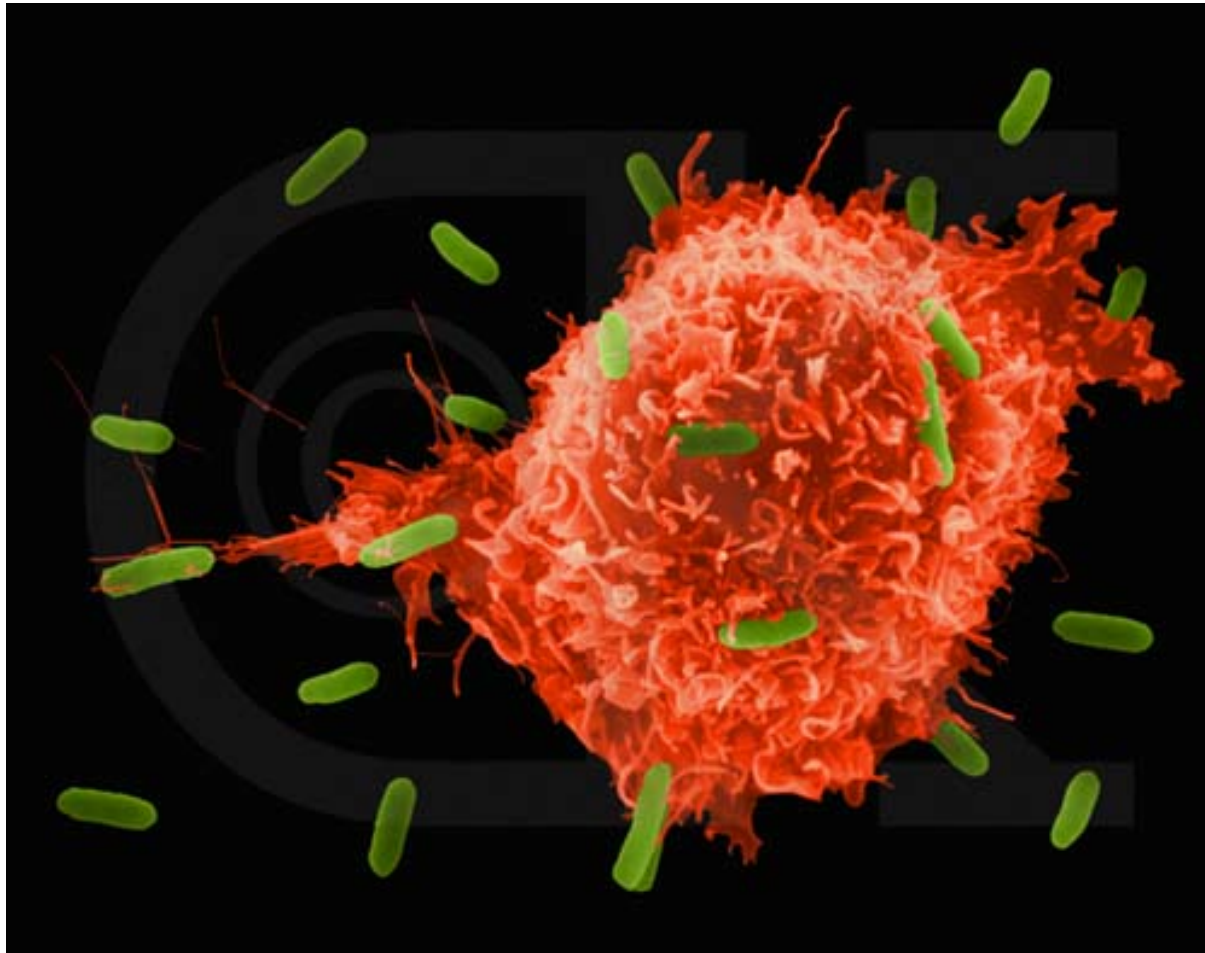


- Act at a distance from promoters to 'Enhance' gene transcription
- Function determined by sequence-specific transcription factors
- Exhibit a distinct epigenetic signature; H3K4me1/2 > H3K4me3, H3K27ac
- ~ 1 million predicted in the human genome

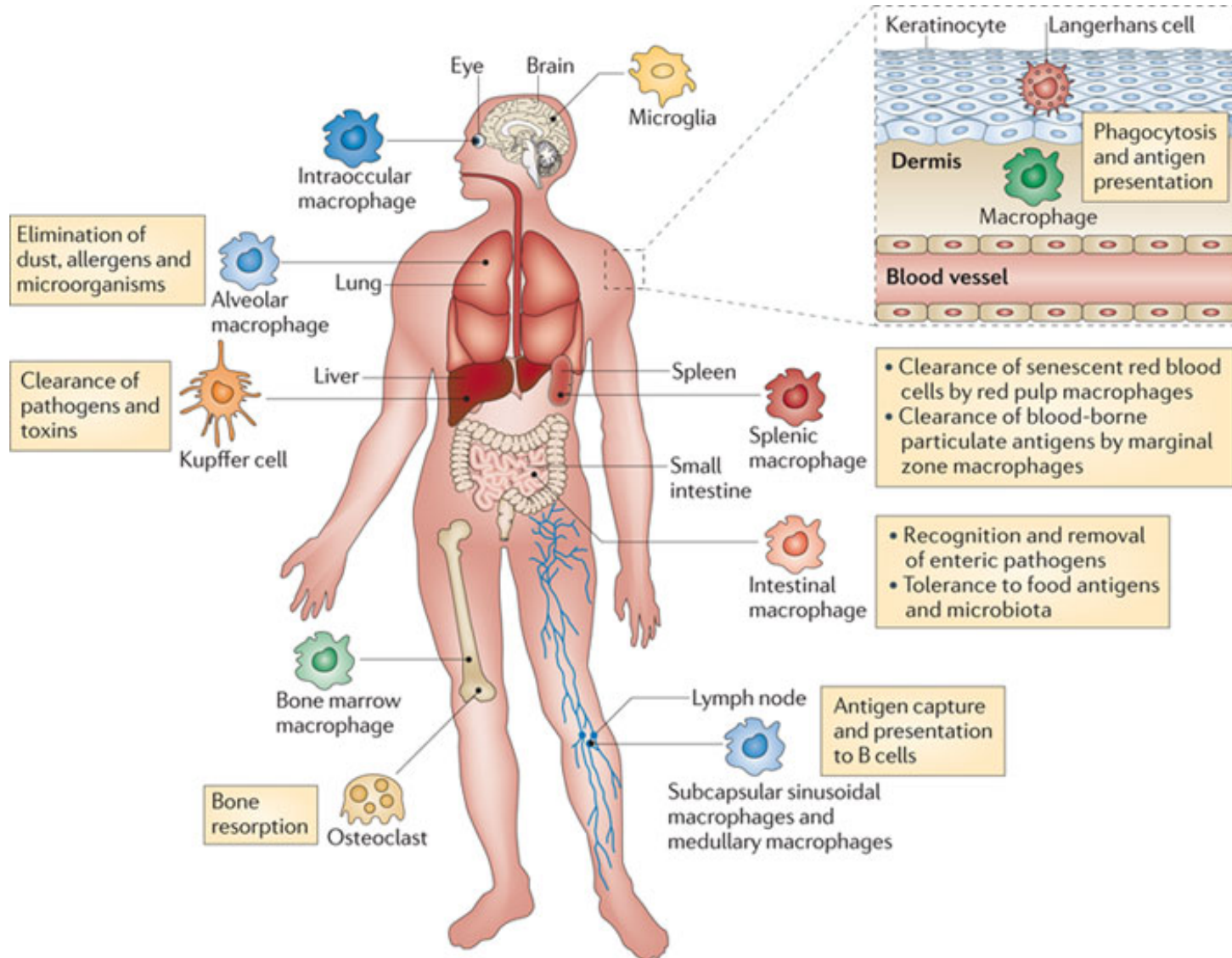
# Cell-specific selection of enhancers



Macrophages play essential roles in the response to infection and injury



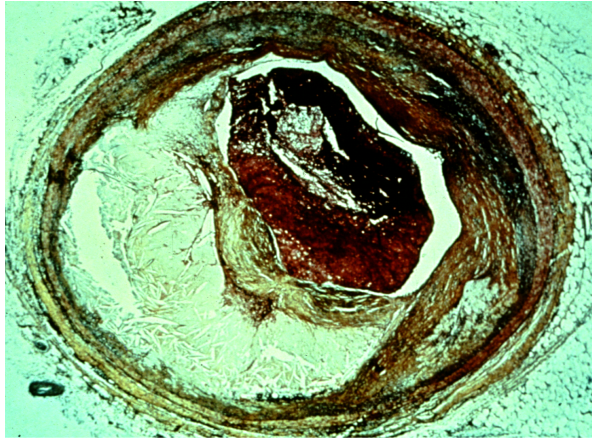
# Specialized homeostatic functions of resident tissue macrophages



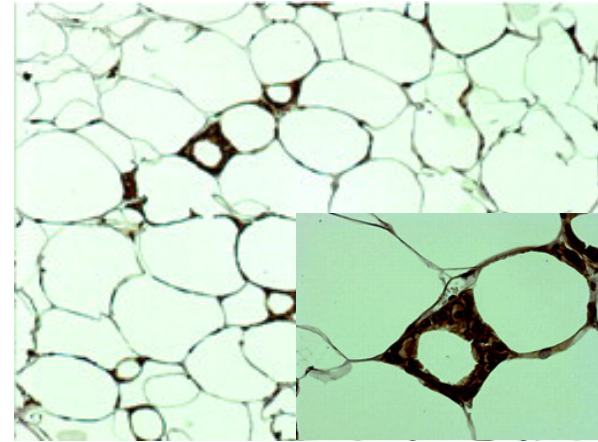


# Roles of macrophages in human disease

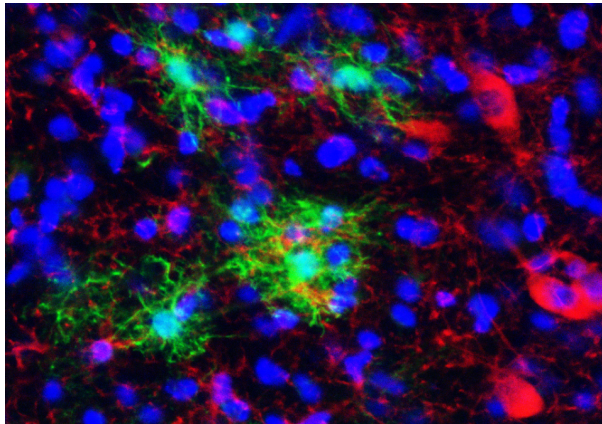
Macrophage foam cells  
in atherosclerosis



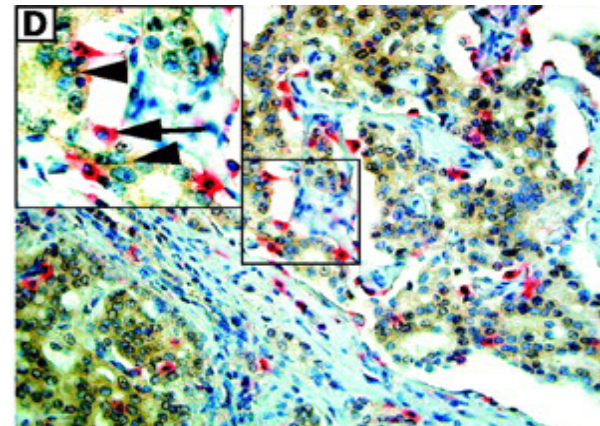
Adipose tissue macrophages  
in insulin resistance



Activated microglia in  
neurodegenerative disease

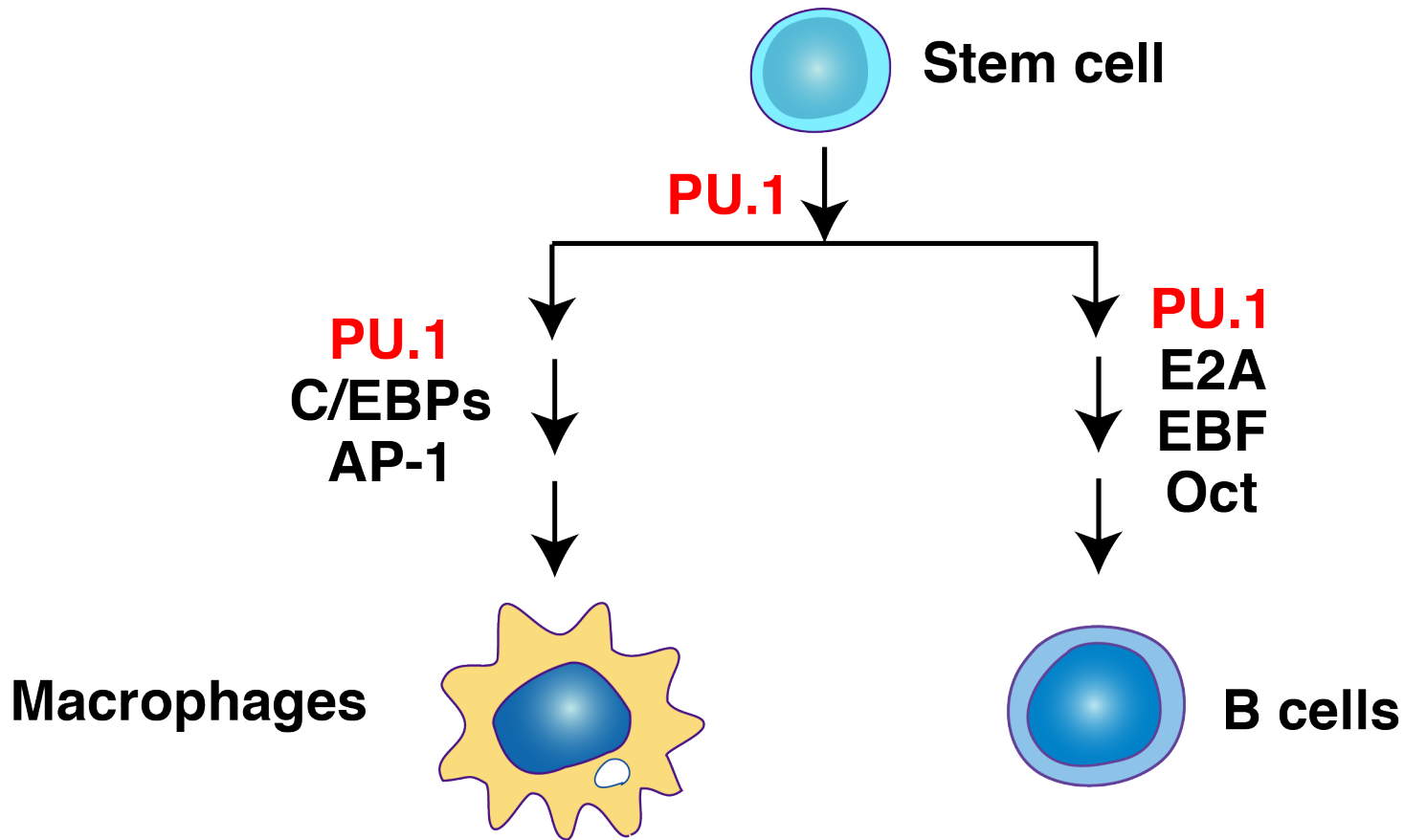


Tumor-associated macrophages  
in cancer





# PU.1 is required for both macrophage and B cell differentiation



# Enriched motifs at macrophage and B-cell-specific PU.1 binding sites

## Macrophage-specific PU.1 binding sites

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	PU.1
	AP1
	C/EBP

## B-cell-specific PU.1 binding sites

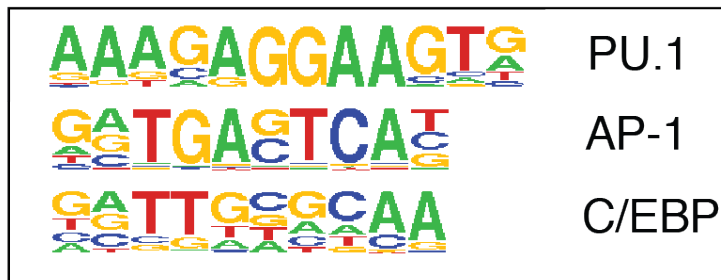
---

	PU.1
	E2A
	EBF
	NFkB
	OCT

# PU.1 and C/EBP $\beta$ occupy the majority of the enhancer-like regions in macrophages

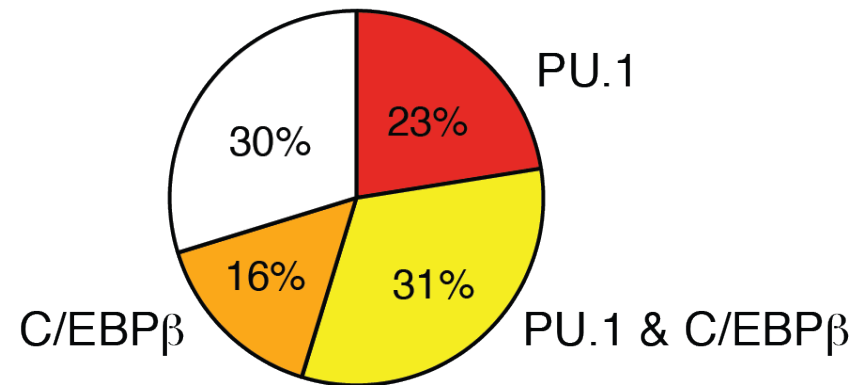
## Macrophage H3K4me1 Peaks

Enriched motifs



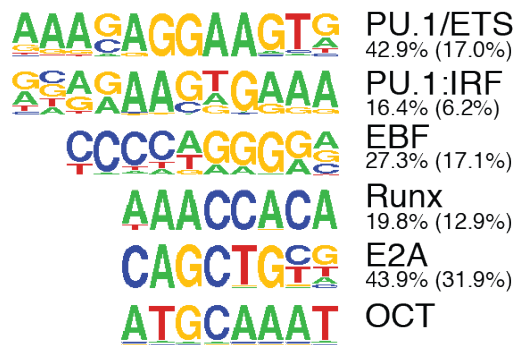
## Macrophage H3K4me1 Peaks

PU.1 and C/EBP $\beta$  ChIP-Seq

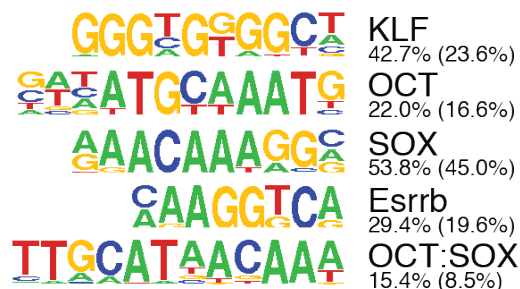


# Motifs for lineage determining TFs are enriched in H3K4me1 marked regions of the genome

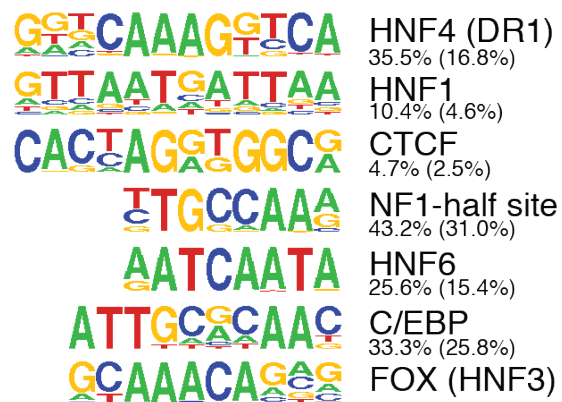
## B cells



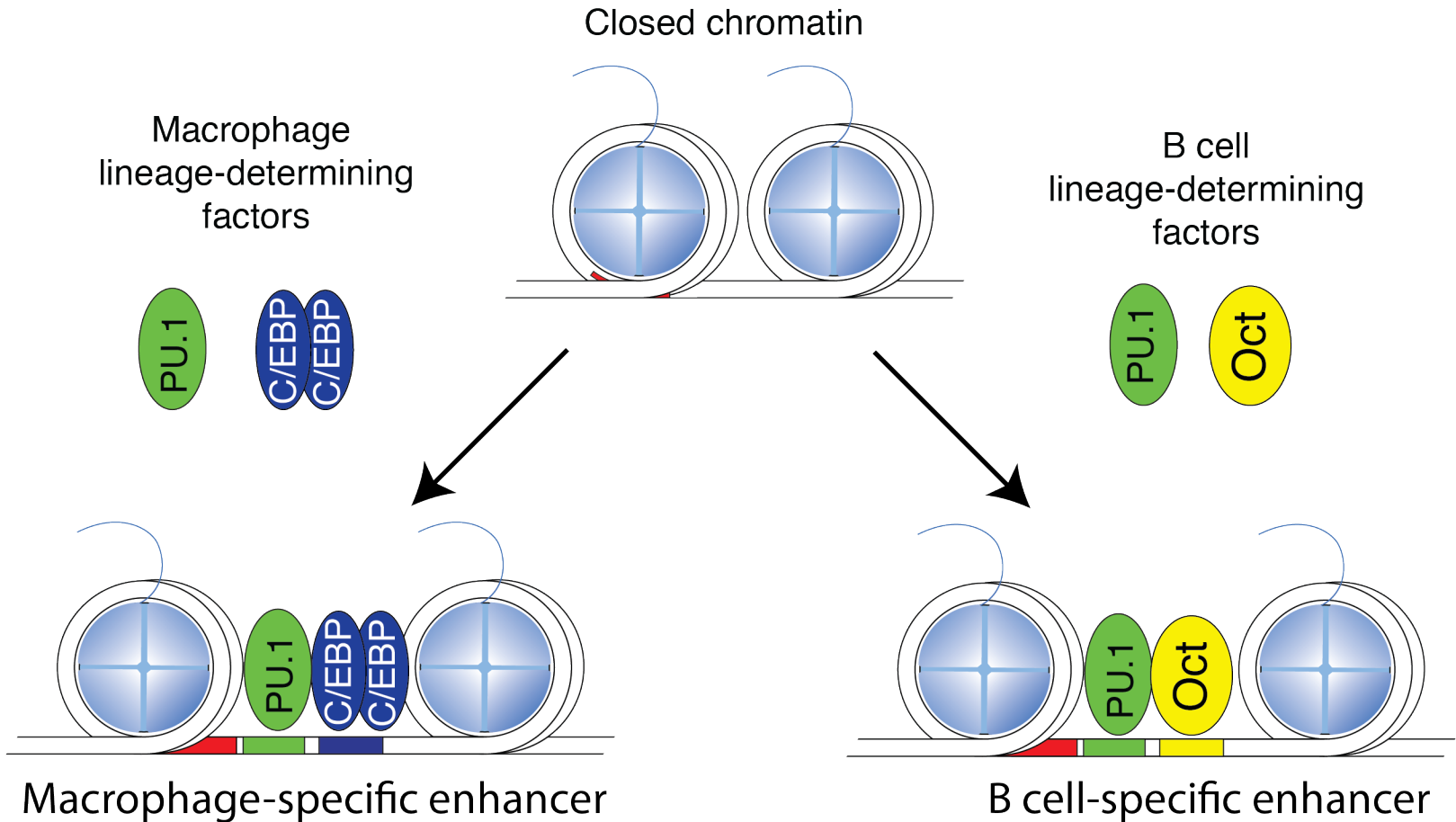
## Embryonic stem cells



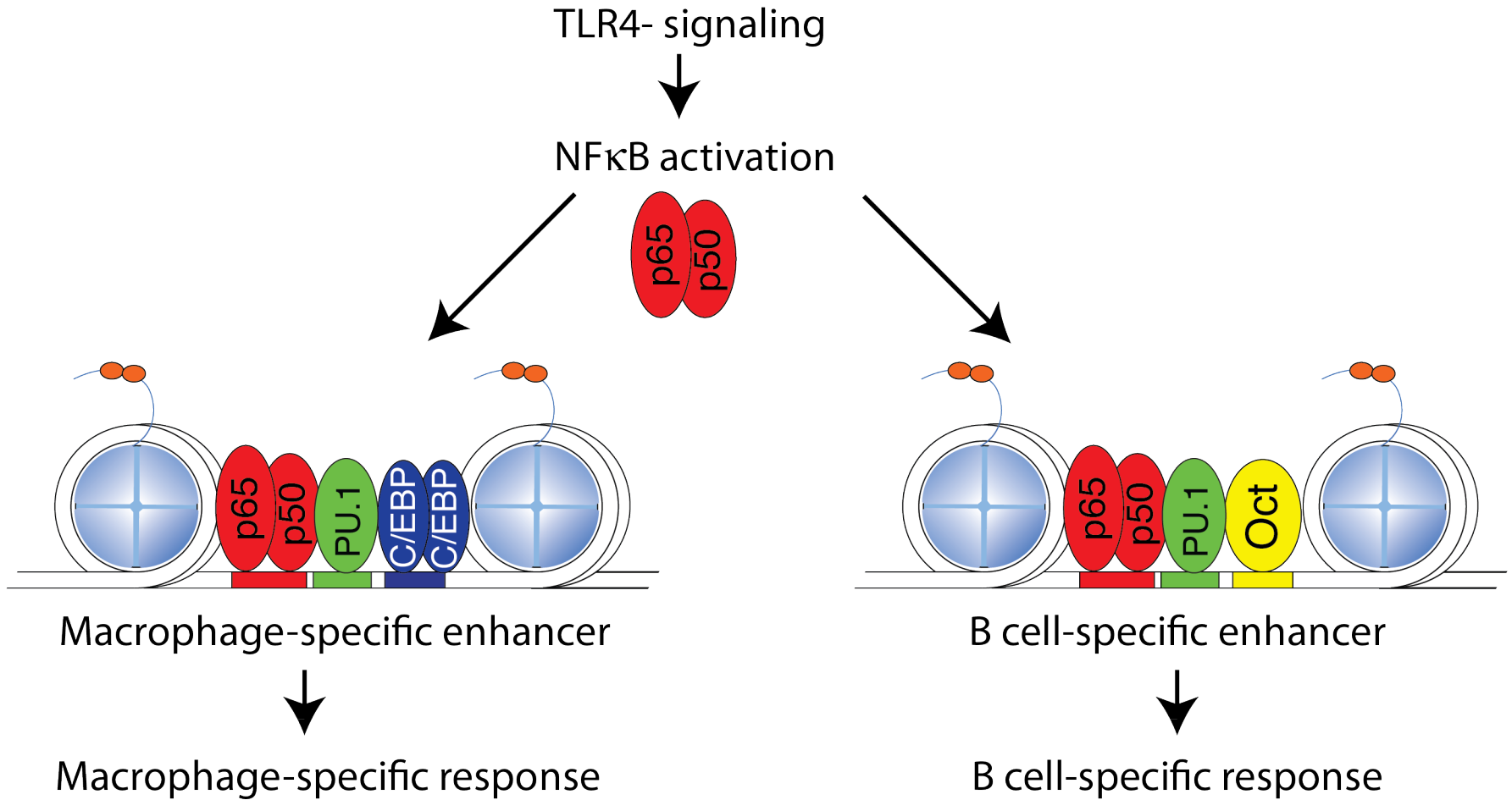
## Liver



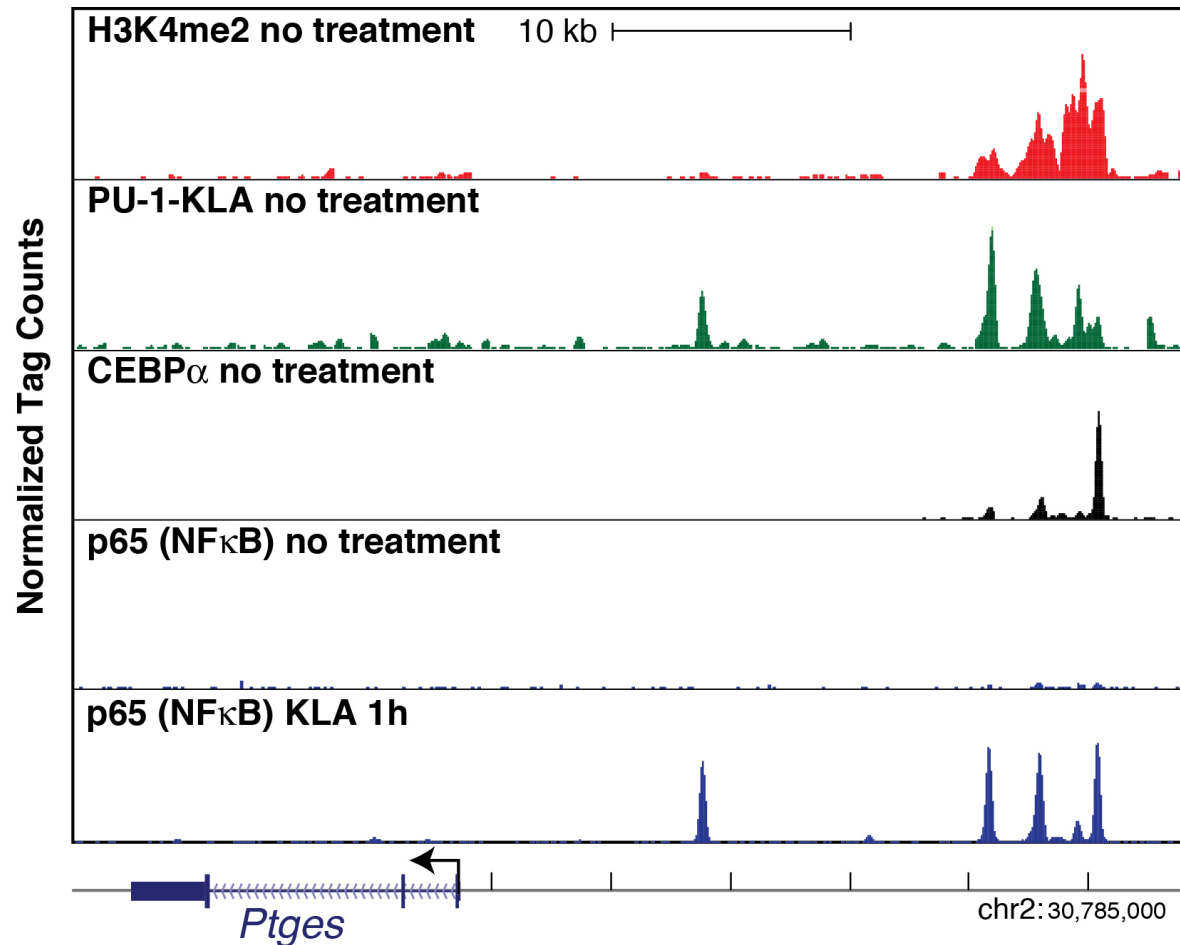
# Selecting macrophage and B cell-specific enhancers



# Selecting macrophage and B cell-specific enhancers

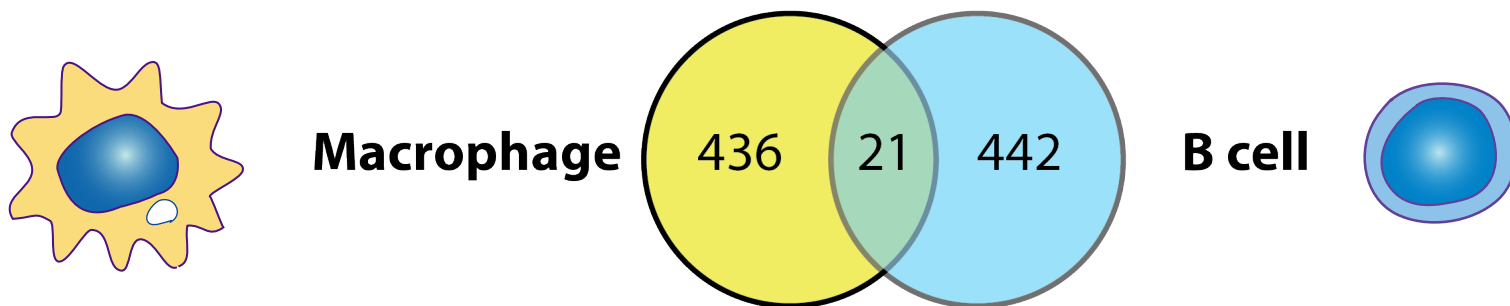


# Binding of p65 (NF $\kappa$ B) to poised enhancer like regions near *Ptges*



# Transcriptional responses to the same signal can be cell type-specific

mRNAs increased > 4-fold 1h after TLR4 ligation



	Macrophage	B cell
GO term	p value	p value
Immune response	1e-36	> 0.05
Chemotaxis	1e-12	> 0.05
Protein folding	> 0.05	1e-10
RNA processing	> 0.05	1e-9
DNA replication	> 0.05	8e-3



# A collaborative – hierarchical model for enhancer selection and activation

## Cell Fate-determining factors (i.e, lineage-determining TFs)

- Expressed in cell-restricted combinations
- Collaborate with each other and additional factors to bind to DNA and initiate nucleosome remodeling
- Establish cellular identity and exhibit reprogramming potential



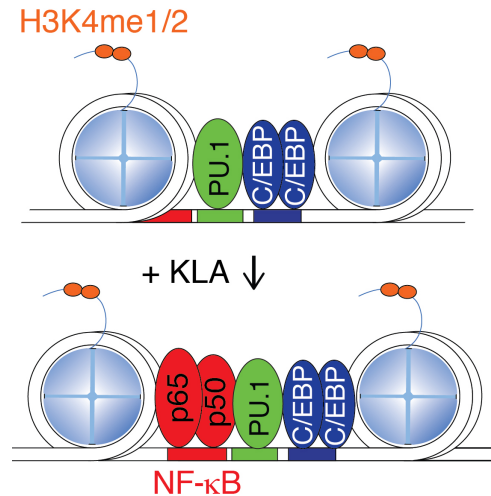
## Cell State-determining factors (i.e., Signal-dependent TFs)

- Broadly expressed
- Primarily localize to pre-existing 'primed'-enhancers
- Confer responsiveness to internal and external signals

# Effects of natural genetic variation support a collaborative/hierarchical model



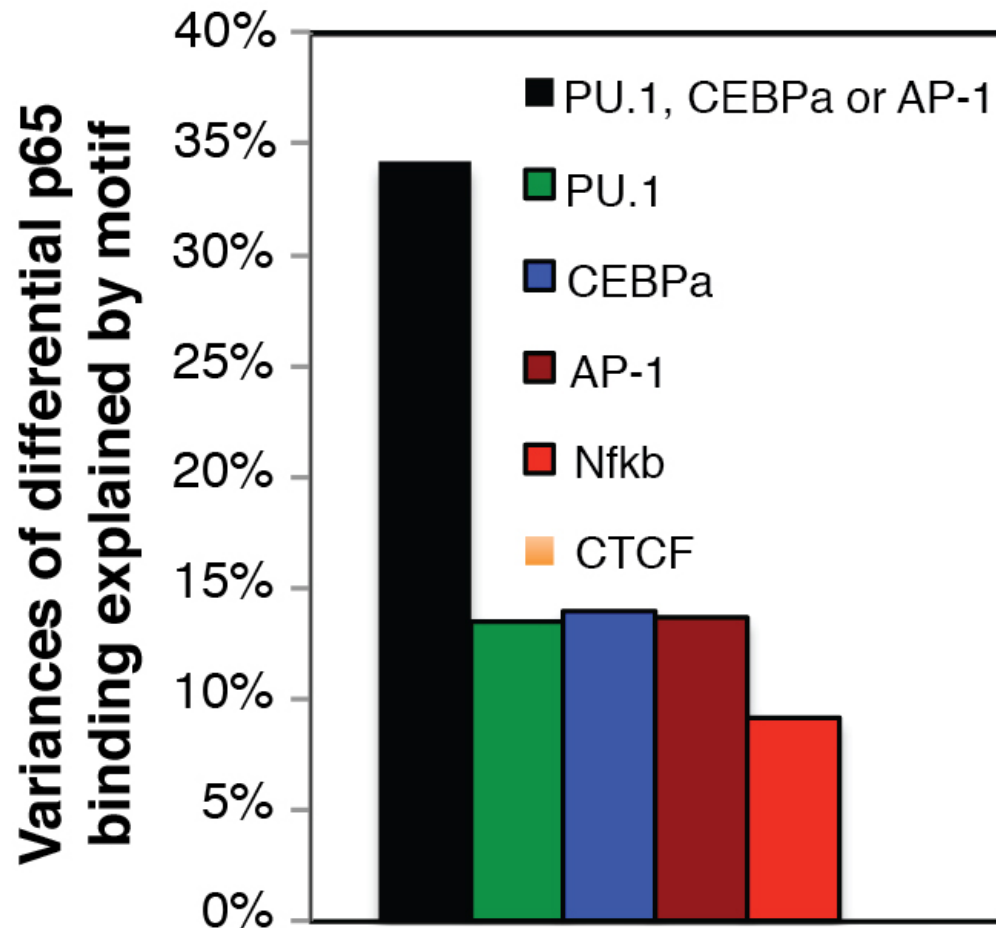
C57BL/6



BALBc ~4m SNPS

- Mutations in PU.1 motifs reduce nearby binding of C/EBP
- Mutations in C/EBP motifs reduce nearby binding of PU.1
- Mutations in NF $\kappa$ B motifs rarely reduce binding of nearby PU.1 or C/EBP
- Mutations in PU.1 or C/EBP motifs frequently reduce binding of NF $\kappa$ B

# Effect of motif mutations on p65 binding



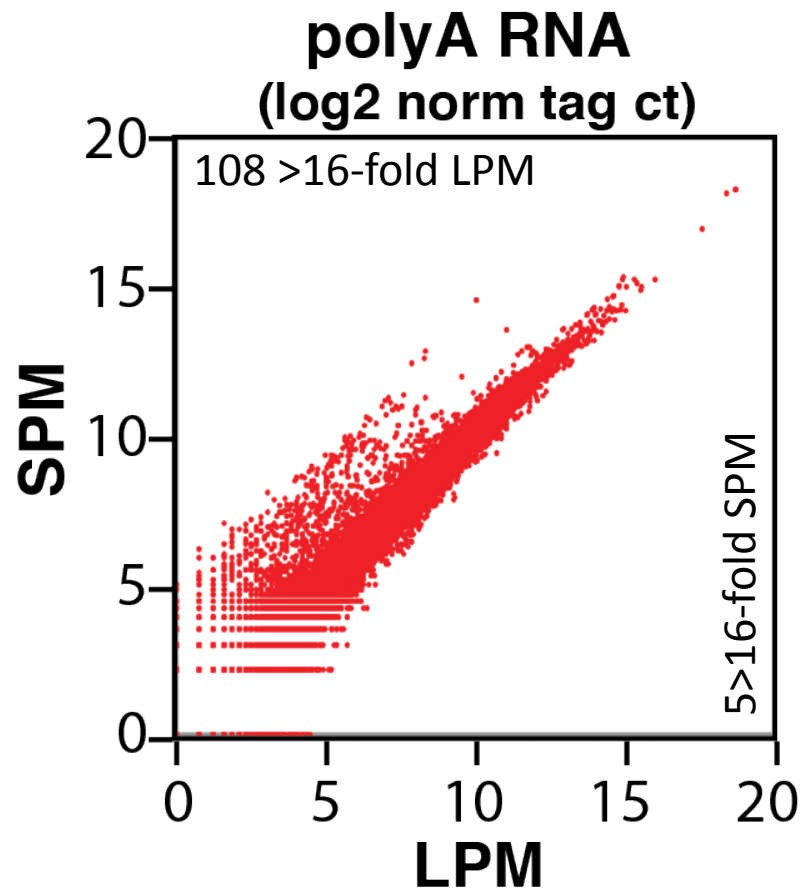
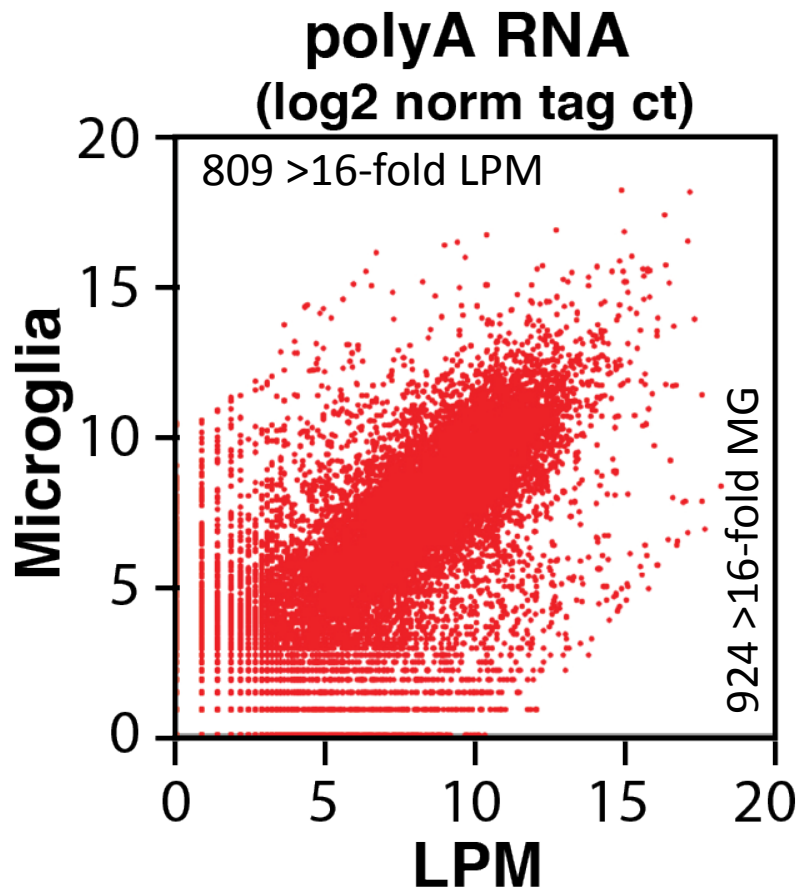
# A collaborative – hierarchical model for enhancer selection and activation

- Supported by gain and loss of function experiments and effects of mutations in transcription factor binding sites (e.g., Heinz et al., Mol Cell, 2010, Heinz et al., Nature 2013, Kaikkonen et al., Mol. Cell, 2013)
- Model is vastly oversimplified and has poor predictive power
- Fails to explain how new enhancers are selected
- Fails to account for functions of the majority of transcription factors expressed in macrophages
- Relevance to in vivo populations of macrophages is unclear

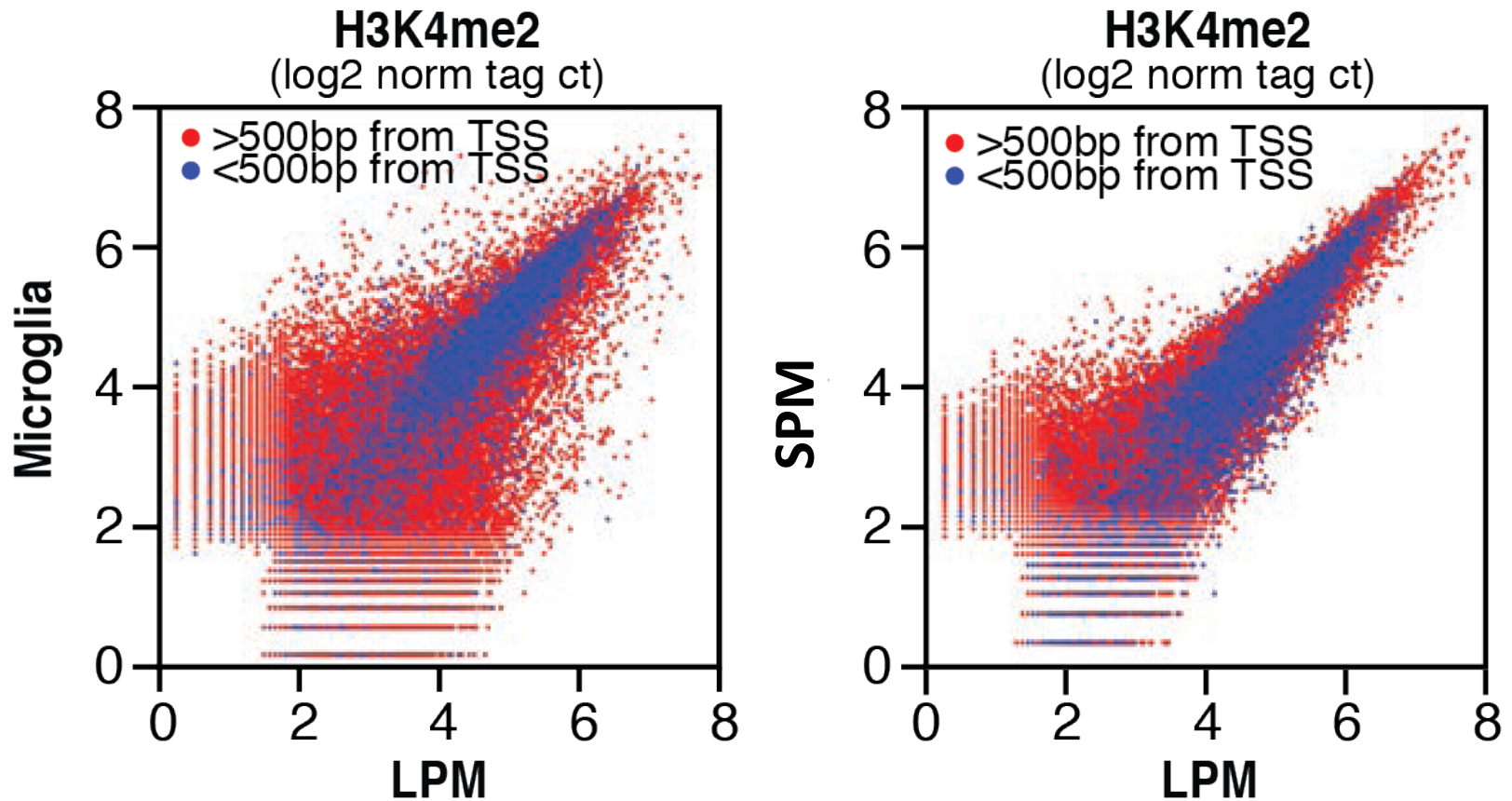
# A comparison of tissue resident macrophage subsets

<b>Cell type</b>	<b>Environment</b>
<b>Microglia (MG)</b>	Brain/TGF $\beta$
<b>Large perit. M<math>\Phi</math> (LPM - MHCII<sup>low</sup>)</b>	Peritoneum
<b>Small perit. M<math>\Phi</math> (SPM - MHCII<sup>med</sup>)</b>	RA

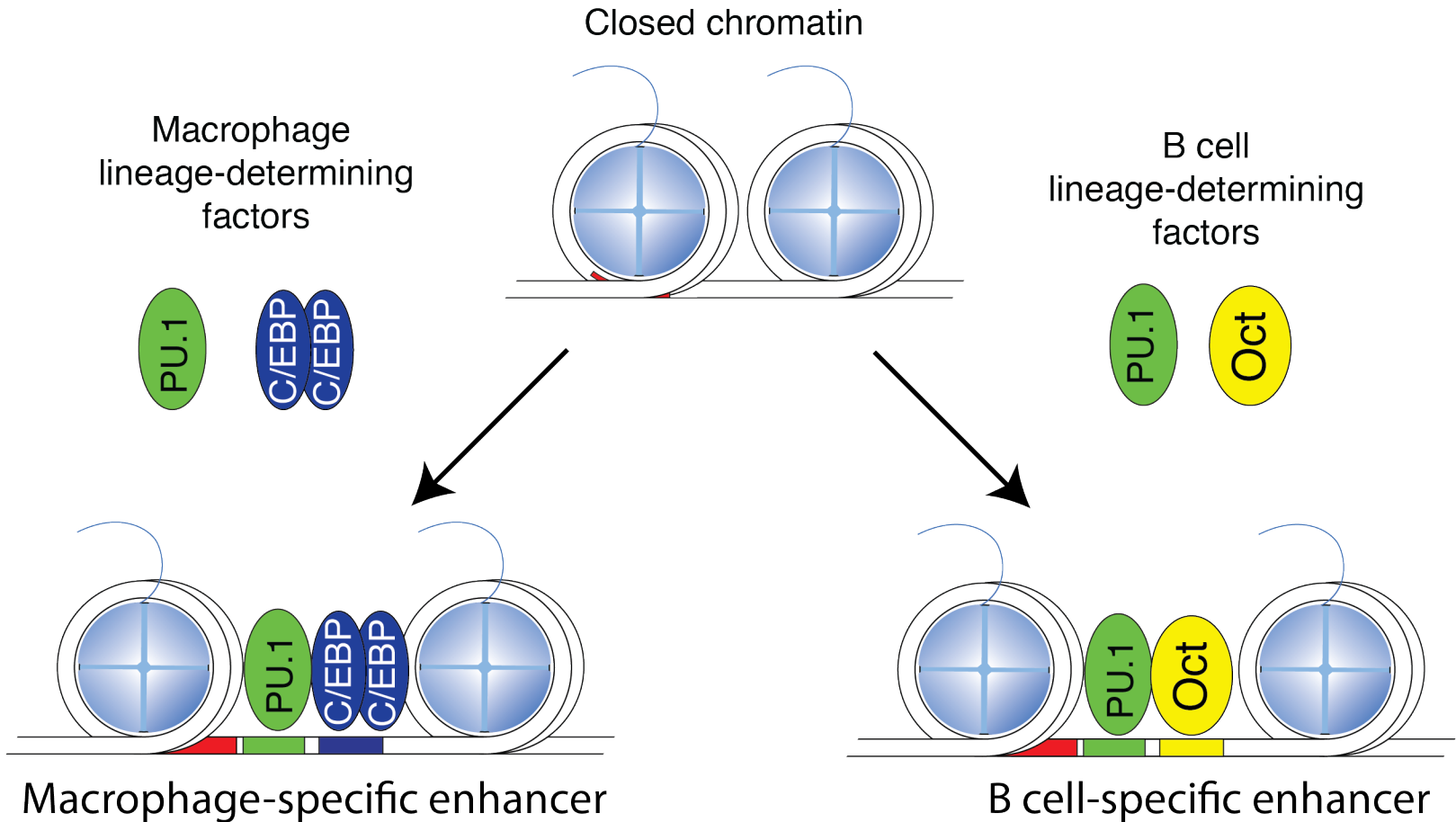
# Macrophage transcriptomes can be highly divergent dependent on tissue of residence



# Macrophage enhancer landscapes differ depending on their source

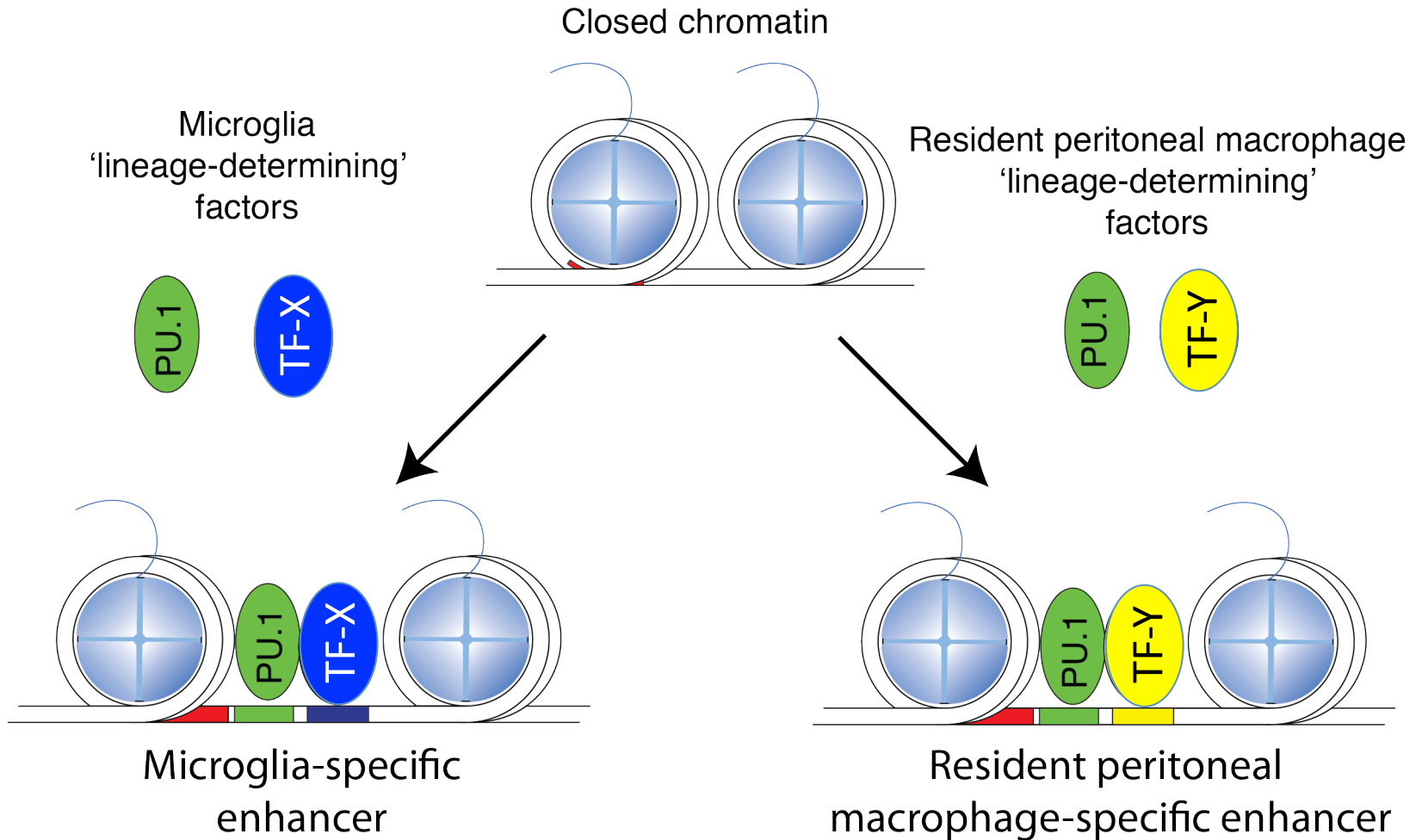


# Selecting macrophage and B cell-specific enhancers



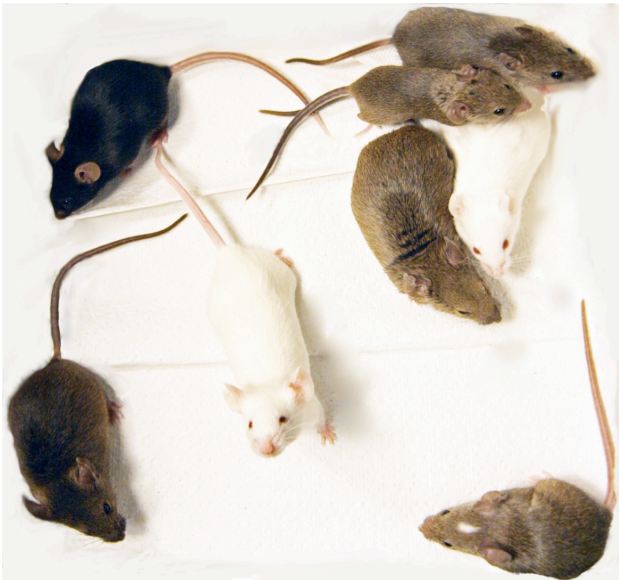


# Selecting microglia- and RPM-specific enhancers



# What TFs collaborate with PU.1 to select microglia and RPM-specific enhancers?

- Strategy :
  - Use the vast natural genetic variation provided by inbred strains of mice as an in vivo ‘mutagenesis screen’



Inbred Strain	SNPs relative to C57BL/6J
BALB/cJ	~4 million
NOD/ShiLt/J	~5 million
SPRET/EiJ	~40 million

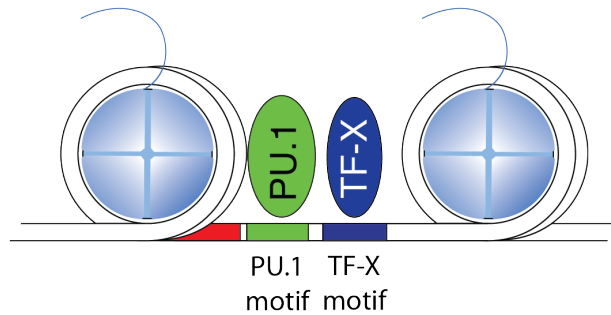
# Use of natural genetic variation to identify motifs for collaborative TFs

Transcription Factor X is a collaborative partner for PU.1 at a subset of PU.1 binding sites in the genome

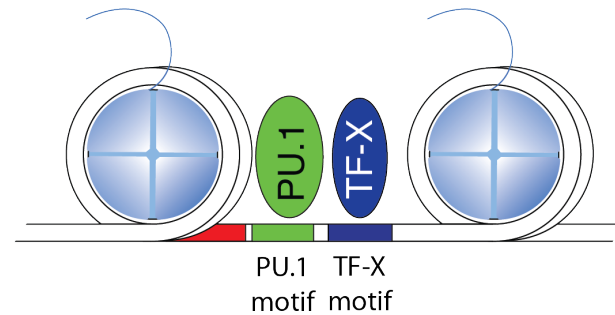
Most genomic locations

Informative genomic locations

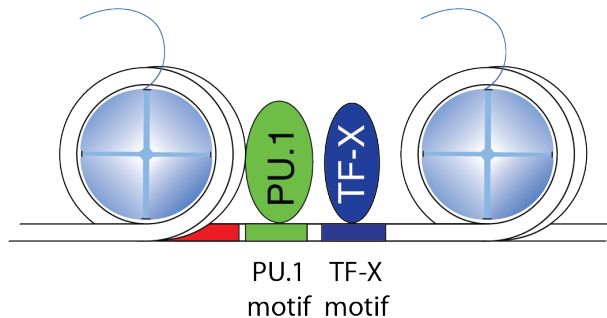
Strain 1



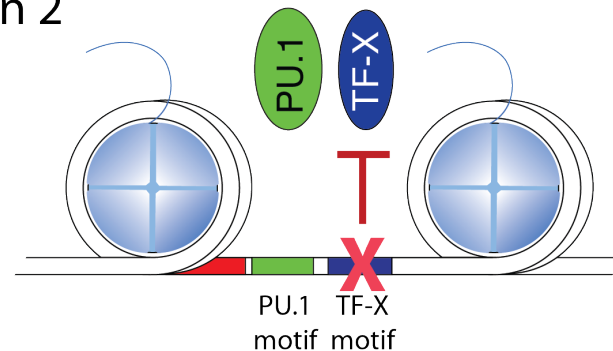
Strain 1



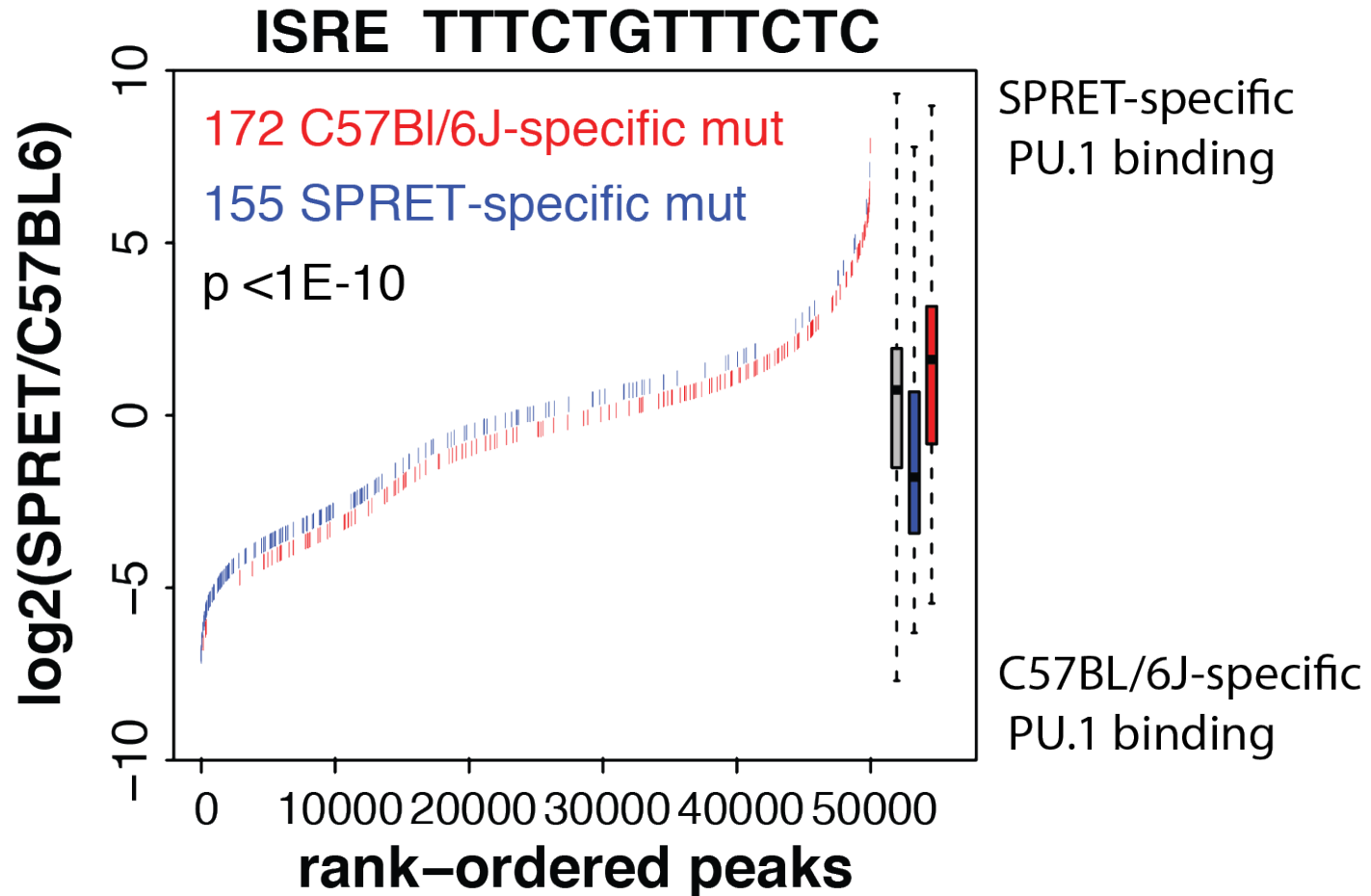
Strain 2



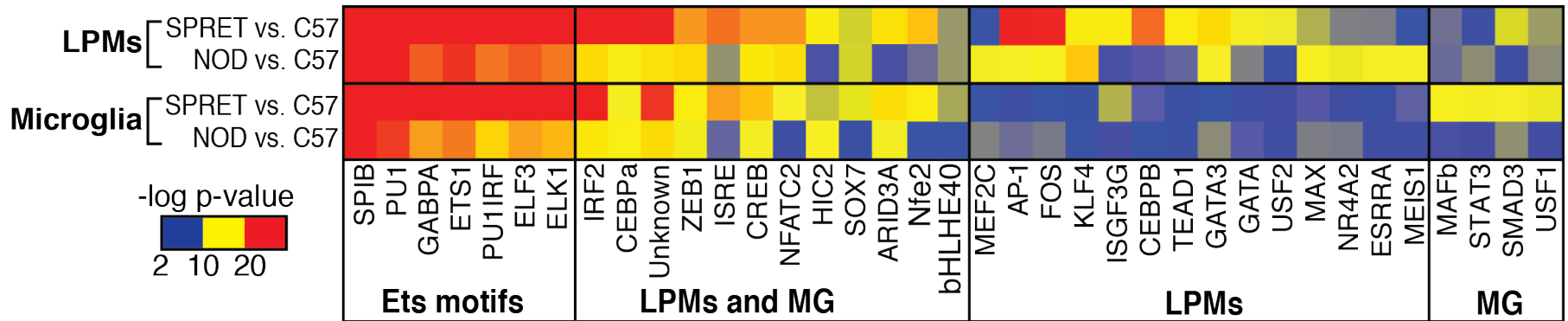
Strain 2



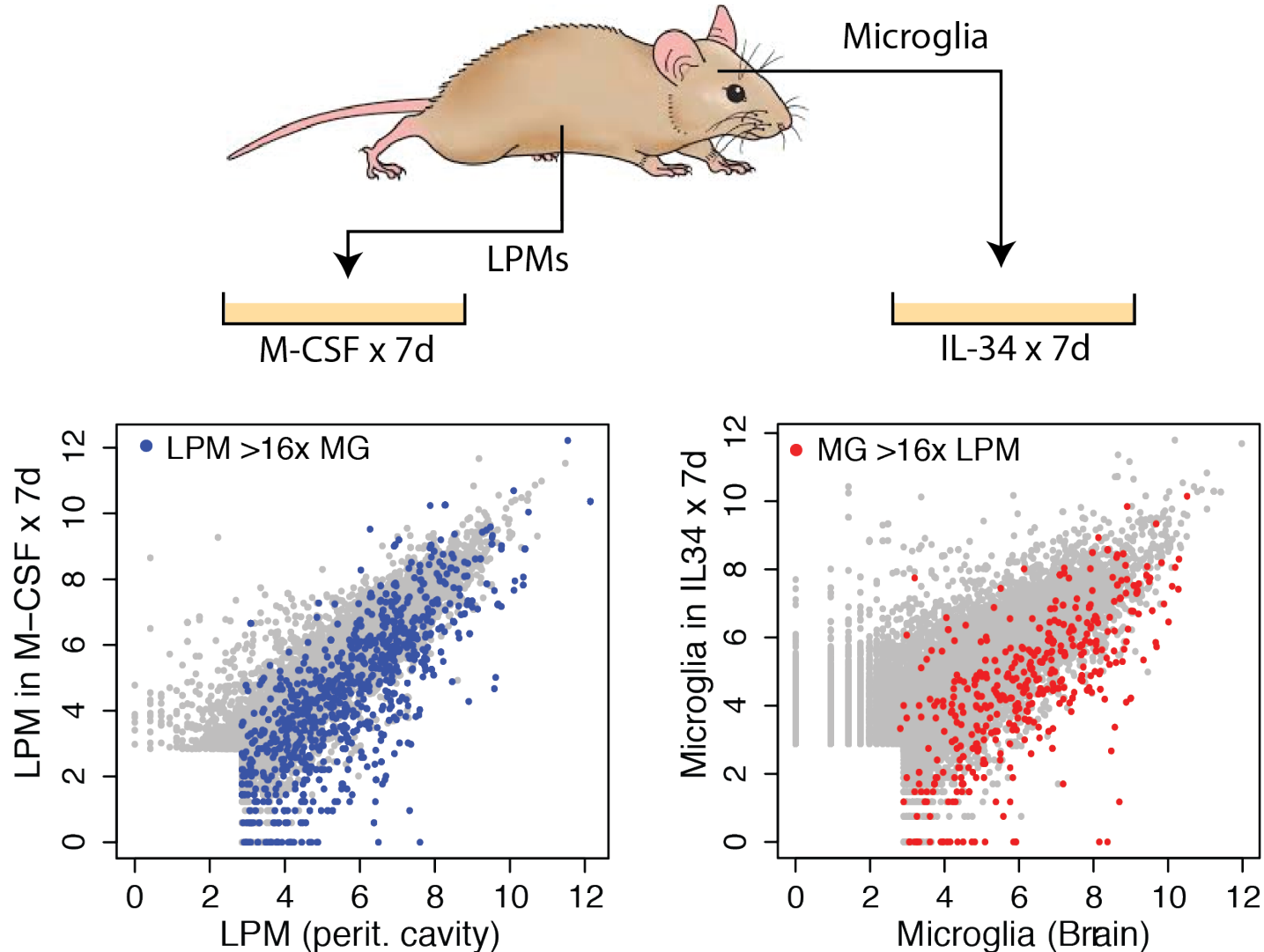
# Mutations in nearby ISRE motifs are highly correlated with strain-specific PU.1 binding



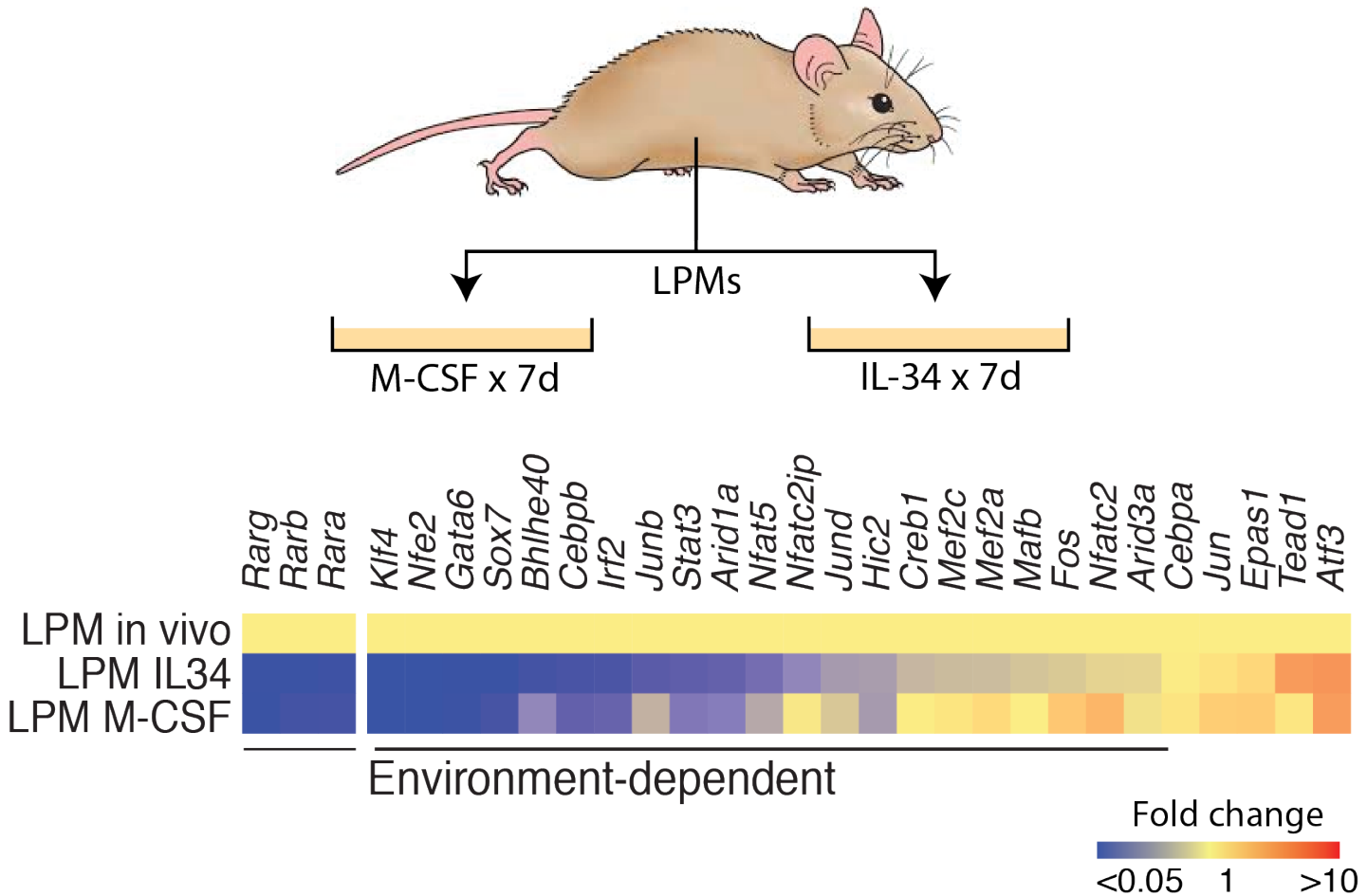
# Discovery of TF motifs associated with common and subset-specific binding of PU.1



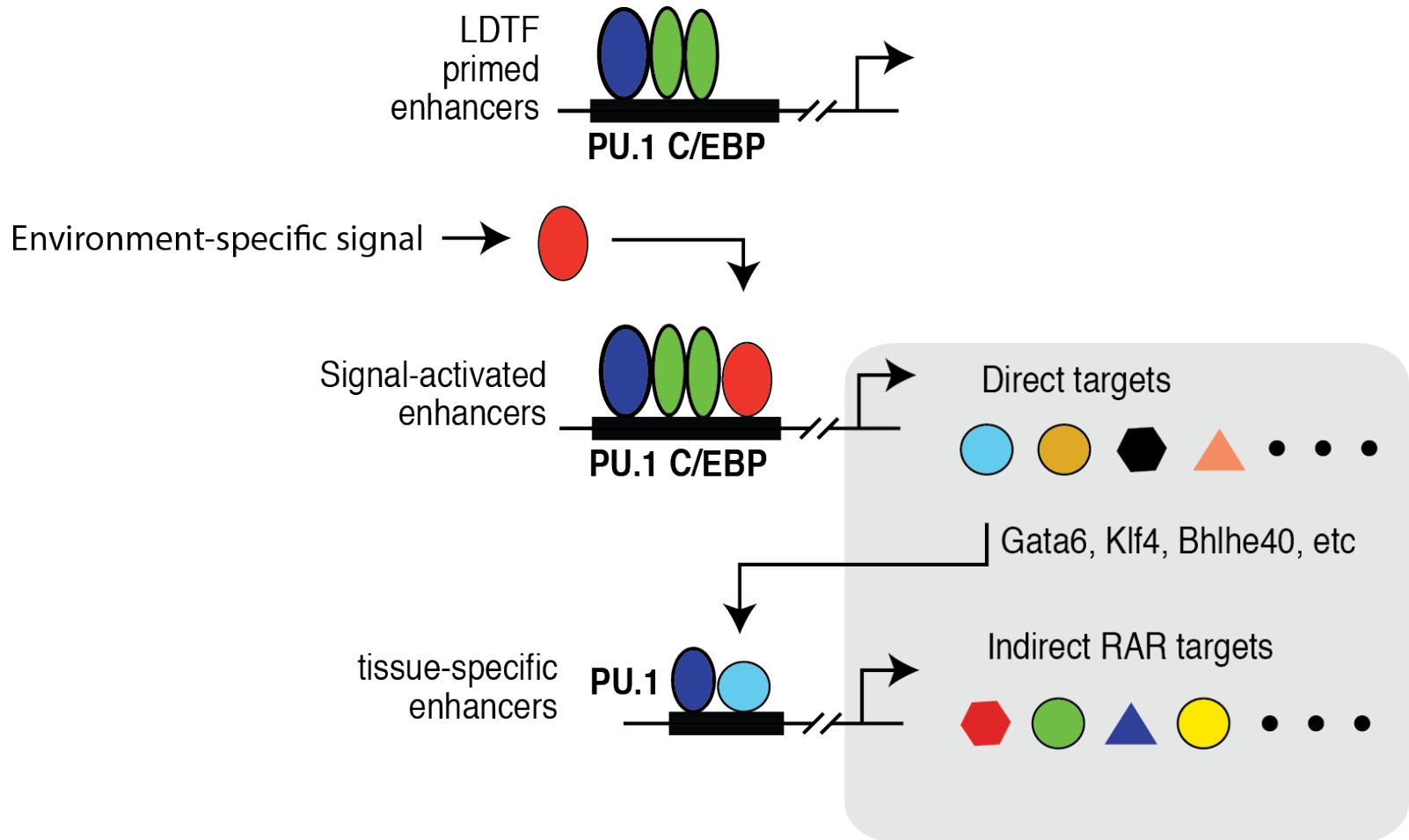
# Macrophage identities require constant environmental signals



# Environment controls expression of TFs that collaborate with PU.1

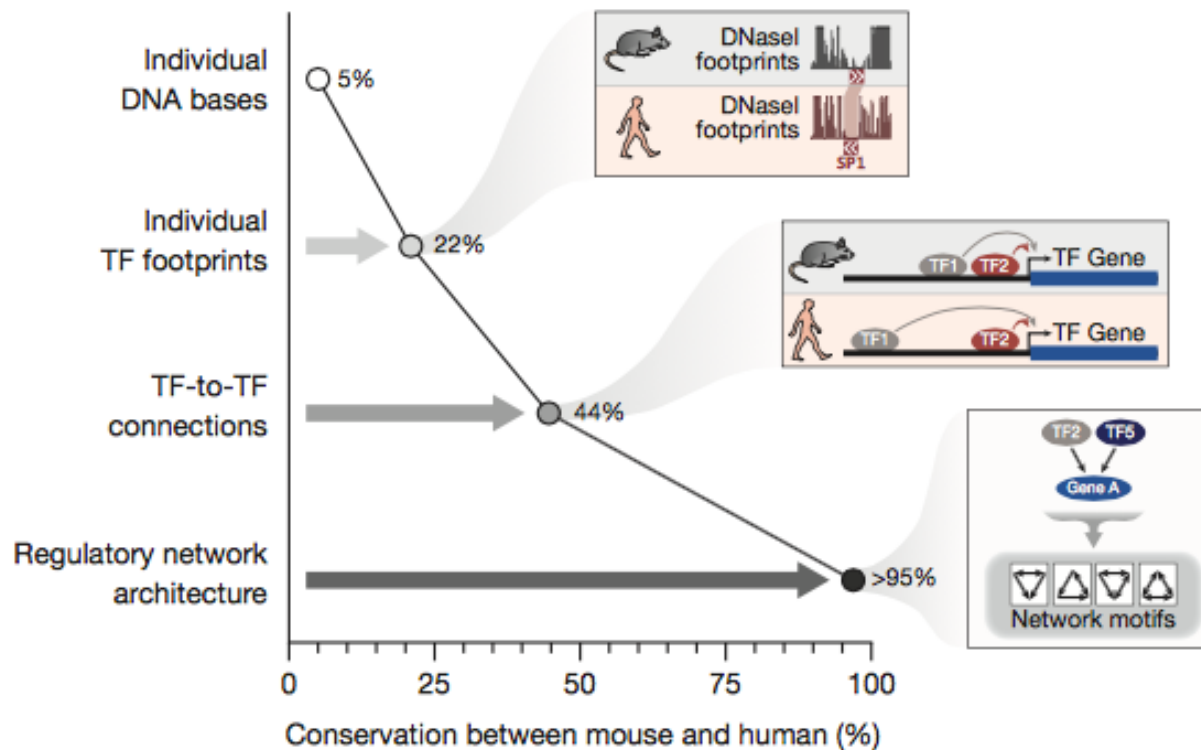


# Environment drives selection and function of enhancers controlling macrophage identities





# Conservation of trans-acting circuitry during mammalian regulatory evolution



# Some take home points

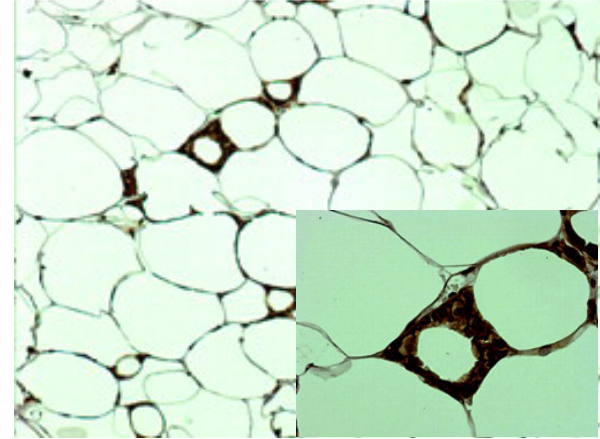
- Knowledge of the enhancer landscape of a cell reveals much about that cell's identity and regulatory potential
- Enhancer landscapes enable prediction of key lineage determining transcription factors and sites of action of signal-dependent factors
- Transcription factor binding maps inform analysis of genetic variation
- Natural genetic variation can be exploited to discover regulatory networks that drive cell-specific gene expression

# Understanding and modifying roles of macrophages in human disease

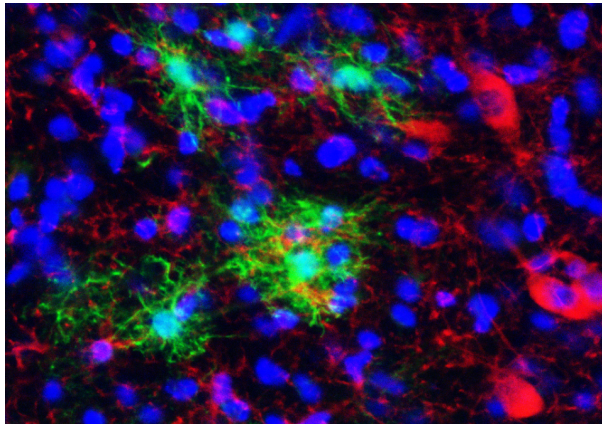
Macrophage foam cells  
in atherosclerosis



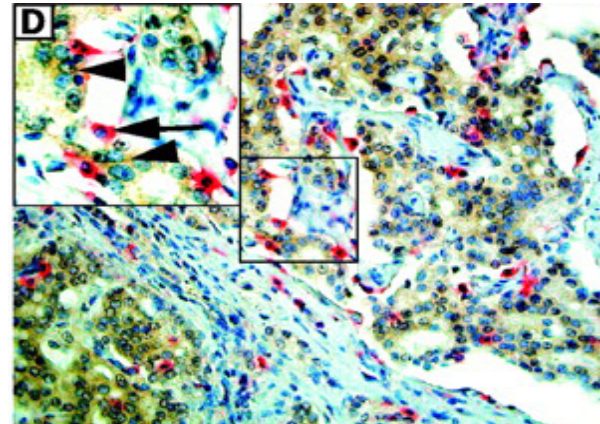
Adipose tissue macrophages  
in insulin resistance



Activated microglia in  
neurodegenerative disease



Tumor-associated macrophages  
in cancer



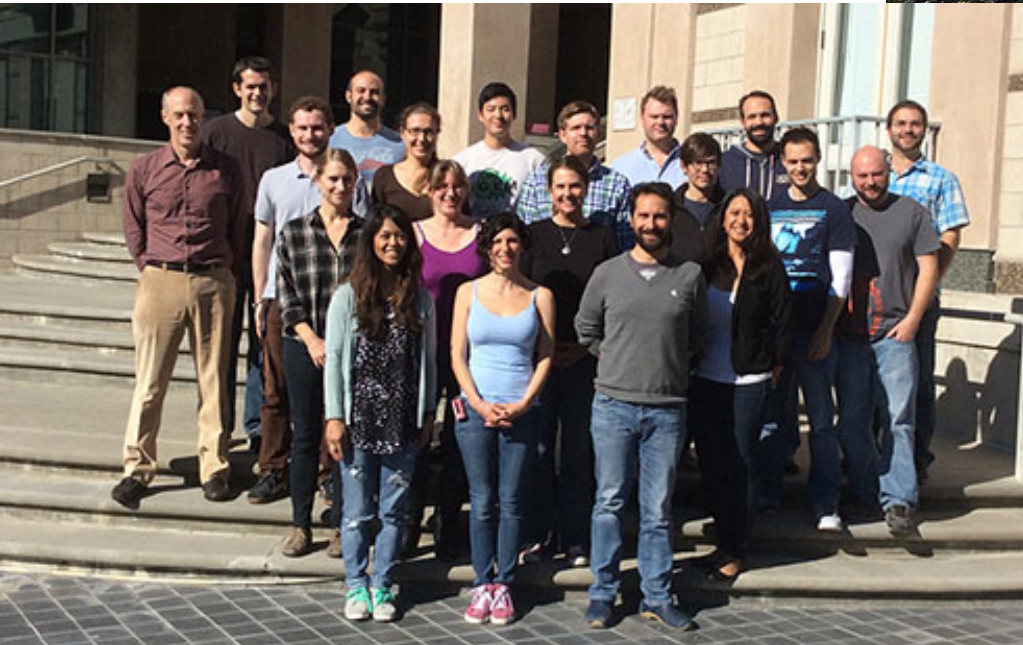
# Going forward

- Improve methodology to define regulatory networks in specific cell types within complex tissues
- Determine effects of cell-autonomous and non-autonomous disease mechanisms on enhancer selection and function
- Consideration of regulatory networks as complex phenotypes for therapeutic modulation



# Thanks!

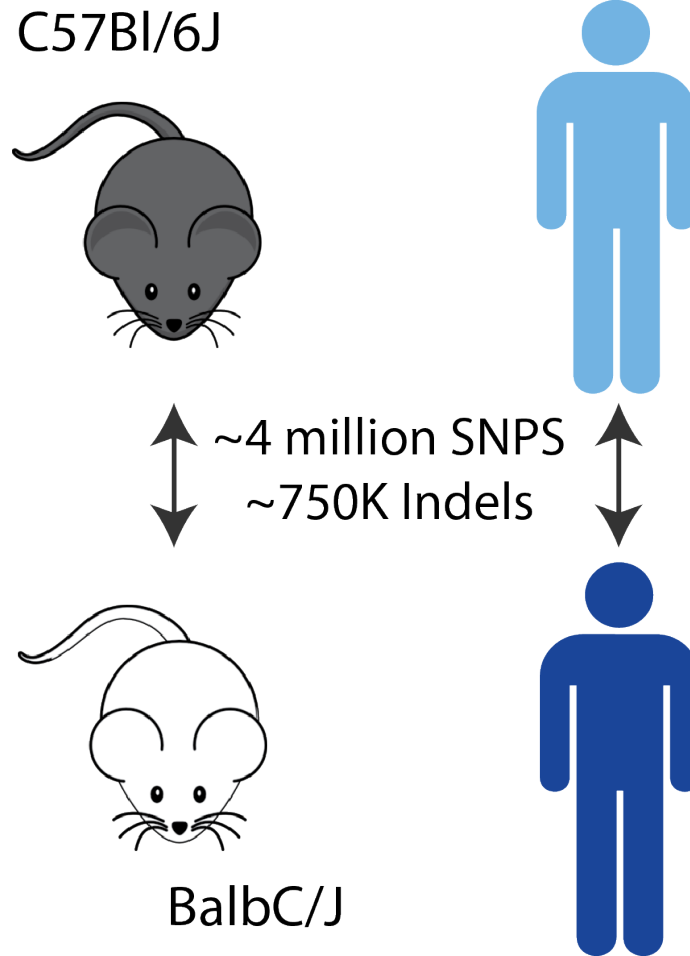
- David Gosselin
- Verena Link
- Casey Romanoski
- Sven Heinz
- Chris Benner
- Dawn Eichenfeld
- Ty Troutman
- Michael Lam



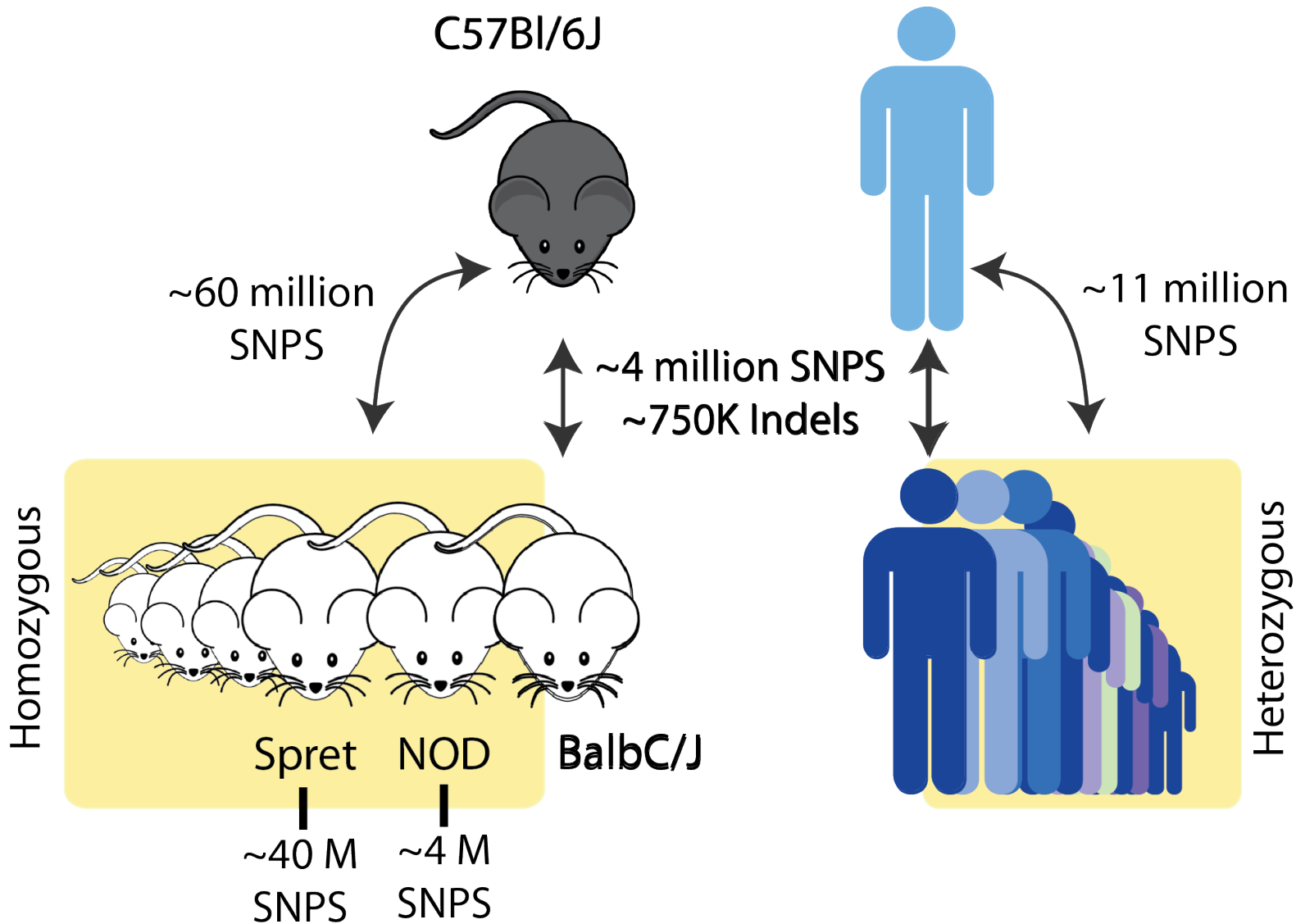
## Kings College

- Frederic Geissmann
- Hanna Gardiner

# Exploiting natural genetic variation to understand enhancer selection and function



# Exploiting natural genetic variation to understand enhancer selection and function



# Gene expression in the same cell type can vary among individuals

